

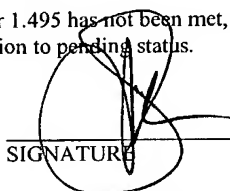
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JC962 U.S. PTO

EXPRESS MAIL MAILING LABEL
EXPRESS MAIL NO.: EL794535160US
DATE OF DEPOSIT: MARCH 15, 2002

JC10 Rec'd PCT/PTO 15 MAR 2002

U.S. APPLICATION NO. (if known, see 37 CFR 1.53) 10/088549		INTERNATIONAL APPLICATION NO. PCT/EP 00/09130		ATTORNEY'S DOCKET NUMBER DEBE:007US	
17. <input checked="" type="checkbox"/> The following fees are submitted: Basic National Fee (37 CFR 1.492(a)(1)-(5)): Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO.....\$1,000.00 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$ 860.00 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$ 710.00 International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4)..... \$ 690.00 International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(2)-(4)..... \$ 100.00 <div style="text-align: right;">ENTER APPROPRIATE BASIC FEE AMOUNT = \$1,000.00</div>				CALCULATIONS PTO USE ONLY	
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				\$.00	
Claims	Number Filed	Number Extra	Rate		
Total Claims	26 - 20 =	6	x \$ 18.00	\$108.00	
Independent Claims	1 - 2 =	0	x \$ 80.00	\$.00	
Multiple dependent claim(s) (if applicable)			+ \$270.00	\$.00	
TOTAL OF ABOVE CALCULATIONS				= \$1,108.00	
Reduction by 1/2 for filing by small entity, if applicable. Applicant is entitled to small entity status pursuant to 37 CFR 1.27.				\$-0.00	
SUBTOTAL				= \$1,108.00	
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).				\$-0.00	
TOTAL NATIONAL FEE				= \$-0.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property)				\$-0.00	
TOTAL FEES ENCLOSED				= \$1,108.00	
				Amount to be refunded:	\$.00
				charged:	\$.00
a. <input type="checkbox"/> A check in the amount of \$_____ cover the above fees is enclosed. b. <input checked="" type="checkbox"/> Please charge Deposit Account No. 50-1212/10201173/SLH in the amount of \$ 1,108.00 to cover the above fees. A duplicate copy of this sheet is enclosed. c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 50-1212/10201173/SLH. A duplicate copy of this sheet is enclosed.					
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.					
SEND ALL CORRESPONDENCE TO: STEVEN L. HIGHLANDER, ESQ. FULBRIGHT & JAWORSKI L.L.P. 600 Congress Avenue, Suite 2400 Austin, Texas 78701 512.474.5201					
			SIGNATURE  Steven L. Highlander NAME 37,642 REGISTRATION NUMBER		

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10/088549

EXPRESS MAIL MAILING LABEL
EXPRESS MAILING NO.: EL794535160US
DATE OF DEPOSIT: MARCH 15, 2002

JC10 Rec'd PCT/PTO 15 MAR 2002

PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:

Erik Neilsen, Savvas Christophoridis, Carol
Murphy, Marino Zerial and Stefano De
Renzis

Group Art Unit: Unknown

Examiner: Unknown

Serial No.: Unknown

Atty. Dkt. No.: DEBE:007US/SLH

Filed: March 14, 2002

For: ASSAY TO DETECT SUBSTANCES
USEFUL FOR THERAPY

PRELIMINARY AMENDMENT

Commissioner for Patents
Washington, D.C. 20231

Dear Sir:

Please consider the following amendments prior to examination of the above-captioned application. It is believed that no fees are occasioned by this filing; however, should any fees under 37 C.F.R. §§ 1.16 to 1.21 be required for any reason, the Commissioner is authorized to deduct said fees from Fulbright & Jaworski L.L.P. Account No.: 50-1212/10201173/DEBE:007US/SLH. Please date stamp and return the enclosed postcard as evidence of receipt.

AMENDMENT

In the Specification

Please insert the following paragraph after line 1 of page 1:

This application claims priority to PCT/EP 00/09130, filed on September 18, 2000, and EP 99 118 385.6, filed March 22, 2001. The entire content of both these applications are incorporated by reference.

In the Claims

Please cancel claims 1-13, without prejudice or disclaimer.

Please add the following claims:

14. (New) A method of screening a substance for use as pharmaceutical agents for the prophylaxis and/or treatment of a proliferative, invasive or cell migration disorder comprising assessing the affect of said substance on a GTPase-GTPase effector interaction.
15. (New) The method of claim 14, wherein the GTPase is of the Rab family.
16. (New) The method of claim 14, wherein the GTPase is Rab4, Rab5, Rab7, Rab11, Rab17, Rab18, or Rab22.
17. (New) The method of claim 14, wherein the disorder is selected from the group consisting of cancer, endometriosis, atherosclerosis, inflammatory disease, allergic disease, infectious diseases, diabetes, Alzheimer's disease, and skin repair disease.
18. (New) The method of claim 17, wherein the infectious disease is AIDS, tuberculosis, pseudotuberculosis, cholera, malaria, gastroenteritis, enteric fever, or typhus.
19. (New) The method of claim 17, wherein the infectious disease is caused by Mycobacterium, Staphylococcus, Toxoplasma, Trypanosoma, Listeria, Salmonella, Legionella, Leishmania, Coxiella, Shigella, Yersinia, Neisseria, Vibrio, or Bartonella

20. (New) The method of claim 17, wherein the infectious disease is caused by an infectious agent that infects cells by the endocytic route and resides intracellularly in phagosomes escaping the cellular killing mechanisms.
21. (New) The method of claim 17, wherein the cancer is a benign tumor, a malignant tumor, a carcinoma, a sarcoma, a melanoma, a leukemia, a glioma, or a neuroblastoma, in particular a lung carcinoma, an osteosarcoma, a lymphoma, a soft tissue sarcoma, a breast carcinoma, a bile cancer, a cervix carcinoma, a cancer of the (small) intestine, of the kidneys, of the cavity of the mouth, a penis carcinoma, an ovary cancer, a stomach cancer, a cancer of the tongue, a brain cancer, a bladder carcinoma, a prostate carcinoma, a liver carcinoma, a carcinoma of the pancreas, and every tumor that invades other tissues and organs distinct from its site of origin.
22. (New) The method of claim 14, wherein the assay is carried out in the presence of a labeled GTPase effector/regulator molecule.
23. (New) The method of claim 22, wherein the label is a fluorescent or radioactive label.
24. (New) The method of claim 14, wherein assessing comprises determining GTPase function.
25. (New) The method of claim 14, wherein assessing comprises determining GTPase interaction with a GTPase effector/regulator molecule.
26. (New) The method of claim 24, wherein GTPase function is determined by measuring GTP/GDP nucleotide exchange, GTP hydrolysis, endosomal motility, and endosomal trafficking.
27. (New) The method of claim 25, wherein a GTPase effector molecule is bound to a substrate.

28. (New) The method of claim 27, wherein the substrate is a chromatographic matrix or a bead.
29. (New) The method of claim 14, wherein the substance comprises one or more of the following functional groups: a halide atom bound to an alkyl, alkenyl, alkynyl or aryl residue, an alcohol group (primary, secondary, tertiary), an ether group, a carbonyl function (aldehyde or ketone), a carboxylic acid group, a carboxylic anhydride group, a carbamoyl group, a haloformyl group, a cyano group, an ester group including a lactone group, a benzyl, phenyl, tolyl, tosyl, sulfonyl group, an amino group (primary, secondary, tertiary), a sterol moiety, an isocyanate, a cyanate, a thioisocyanate, a thiocyanate, a carbamate, an azide, a diazo group, or a quinone group.
30. (New) The method of claim 14, wherein the substance is an organometallic compound, a β -hydroxy carboxylic acid, an inorganic acid or complex such as a metallocene, a nucleic acid.
31. (New) The method of claim 30, wherein the antibody is a polyclonal or monoclonal antibody, or a fragment thereof, a humanised or human antibody, an inhibitory or stimulatory antibody.
32. (New) The method of claim 14, wherein the substance is a protein or peptide.
33. (New) The method of claim 32, wherein the protein is a cytokine, a hormone, or an antibody.
34. (New) The method of claim 32, wherein the peptide is an oligopeptide comprising up to 20 amino acid residues
35. (New) The method of claim 34, wherein the oligopeptide is about 8, about 10 or about 12 amino acid residues in length.

36. (New) The method of claim 14, wherein the substance is a nucleic acid.
37. (New) The method of claim 36, wherein the nucleic acid is genomic DNA, cDNA, or mRNA, an oligonucleotide, or an oligoribonucleotide, wherein said nucleic acid encodes all or a fragment of a proteinaceous GTPase effector.
38. (New) The method of claim 37, wherein the encoding sequence is SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, or and 15.
39. (New) The method of claim 37, wherein the nucleic acid further comprises a gene therapy vector.

REMARKS

Should the examiner have any questions regarding the content of this preliminary amendment, a telephone call to the undersigned is invited.

Respectfully submitted,



Steven L. Highlander

Reg. No. 37,642

Attorney for Erik Nielson, Savvas
Chritophoridis, Carol Murphy,
Marino Zerial and Stefano De Renzis

FULBRIGHT & JAWORSKI L.L.P.
600 Congress Avenue, Suite 2400
Austin, Texas 78701
(512) 536-3184

Date: March 15, 2002

MARKED UP COPY OF CLAIMS

1. (Canceled) Use of an effector of a GTPase as a target in an *in vitro* or *in vivo* assay to detect substances useful as pharmaceutical agents for the prophylaxis and/or treatment of cancer and other proliferative, invasive or cell migration disorders such as endometriosis, atherosclerosis, inflammatory and allergic diseases, infectious diseases, diabetes, Alzheimer's disease and skin repair diseases such as psoriasis.
2. (Canceled) Use of claim 1, wherein the GTPase is of the Rab family.
3. (Canceled) Use of claim 2, wherein the GTPase is any of Rab4, Rab5, Rab7, Rab11, Rab17, Rab18, and Rab22.
4. (Canceled) Use of any of the preceding claims, wherein the infectious disease is AIDS, tuberculosis, pseudotuberculosis, cholera, malaria, gastroenteritis, enteric fever, typhus, those diseases caused by pathogens (bacteria or organisms) such as Mycobacterium, Staphylococcus, Toxoplasma, Trypanosoma, Listeria, Salmonella, Legionella, Leishmania, Coxiella, Shigella, Yersinia, Neisseria, Vibrio, Bartonella, or any other infectious disease caused by an infectious agent that infects cells by the endocytic route and resides intracellularly in phagosomes escaping the cellular killing mechanisms.
5. (Canceled) Use of any of claims 1 to 3, wherein the cancer is a benign tumor, a malignant tumor, a carcinoma, a sarcoma, a melanoma, a leukemia, a glioma, or a neuroblastoma, in particular a lung carcinoma, an osteosarcoma, a lymphoma, a soft tissue sarcoma, a breast carcinoma, a bile cancer, a cervix carcinoma, a cancer of the (small) intestine, of the kidneys, of the cavity of the mouth, a penis carcinoma, an ovary cancer, a stomach cancer, a cancer of the tongue, a brain cancer, a bladder carcinoma, a prostate carcinoma, a liver carcinoma, a carcinoma of the pancreas, and every tumor that invades other tissues and organs distinct from its site of origin.
6. (Canceled) Use of any of the preceding claims, wherein the assay is carried out in the presence of one or more GTPase effector/regulator molecule(s) which is/are either native

- 25144966.1

ID NO: 1, 3, 5, 7, 9, 11, 13, and 15, or gene therapy vectors derived from the aforementioned GTPase effector gene sequences.

11. (Canceled) A kit useful for carrying out the assay of any of the preceding claims, the kit comprising in a suitable means at least one GTPase effector/regulator molecule which is either native and biochemically purified from a vertebrate, or recombinant and biochemically purified from bacterial cultures, from yeast cultures, or from other cultured eukaryotic cells, in either case labeled by a covalent modification or by radioactivity suitable for the use in the assay.
12. (Canceled) The kit of claim 11, wherein the kit further comprises at least one type of GTPase and/or endosomal membrane fractions fluorescently labeled or labeled by any other modification that allows its detection, and/or cytosolic extracts, and/or an ATP-regenerating system and/or a number of chemicals to be tested for their suitability as a drug effective against any of the diseases depicted in claims 1, 4, and 5.
13. (Canceled) The kit of claim 12, wherein the at least one GTPase is/are one or more of the GTPases Rab4, Rab5, Rab7, Rab11, Rab17, Rab18, and Rab22.

Please add the following claims:

14. (New) A method of screening a substance for use as pharmaceutical agents for the prophylaxis and/or treatment of a proliferative, invasive or cell migration disorder comprising assessing the affect of said substance on a GTPase-GTPase effector interaction.
15. (New) The method of claim 14, wherein the GTPase is of the Rab family.
16. (New) The method of claim 14, wherein the GTPase is Rab4, Rab5, Rab7, Rab11, Rab17, Rab18, or Rab22.

17. (New) The method of claim 14, wherein the disorder is selected from the group consisting of cancer, endometriosis, atherosclerosis, inflammatory disease, allergic disease, infectious diseases, diabetes, Alzheimer's disease, and skin repair disease.
18. (New) The method of claim 17, wherein the infectious disease is AIDS, tuberculosis, pseudotuberculosis, cholera, malaria, gastroenteritis, enteric fever, or typhus.
19. (New) The method of claim 17, wherein the infectious disease is caused by Mycobacterium, Staphylococcus, Toxoplasma, Trypanosoma, Listeria, Salmonella, Legionella, Leishmania, Coxiella, Shigella, Yersinia, Neisseria, Vibrio, or Bartonella
20. (New) The method of claim 17, wherein the infectious disease is caused by an infectious agent that infects cells by the endocytic route and resides intracellularly in phagosomes escaping the cellular killing mechanisms.
21. (New) The method of claim 17, wherein the cancer is a benign tumor, a malignant tumor, a carcinoma, a sarcoma, a melanoma, a leukemia, a glioma, or a neuroblastoma, in particular a lung carcinoma, an osteosarcoma, a lymphoma, a soft tissue sarcoma, a breast carcinoma, a bile cancer, a cervix carcinoma, a cancer of the (small) intestine, of the kidneys, of the cavity of the mouth, a penis carcinoma, an ovary cancer, a stomach cancer, a cancer of the tongue, a brain cancer, a bladder carcinoma, a prostate carcinoma, a liver carcinoma, a carcinoma of the pancreas, and every tumor that invades other tissues and organs distinct from its site of origin.
22. (New) The method of claim 14, wherein the assay is carried out in the presence of a labeled GTPase effector/regulator molecule.
23. (New) The method of claim 22, wherein the label is a fluorescent or radioactive label.
24. (New) The method of claim 14, wherein assessing comprises determining GTPase function.

25. (New) The method of claim 14, wherein assessing comprises determining GTPase interaction with a GTPase effector/regulator molecule.
26. (New) The method of claim 24, wherein GTPase function is determined by measuring GTP/GDP nucleotide exchange, GTP hydrolysis, endosomal motility, and endosomal trafficking.
27. (New) The method of claim 25, wherein a GTPase effector molecule is bound to a substrate.
28. (New) The method of claim 27, wherein the substrate is a chromatographic matrix or a bead.
29. (New) The method of claim 14, wherein the substance comprises one or more of the following functional groups: a halide atom bound to an alkyl, alkenyl, alkynyl or aryl residue, an alcohol group (primary, secondary, tertiary), an ether group, a carbonyl function (aldehyde or ketone), a carboxylic acid group, a carboxylic anhydride group, a carbamoyl group, a haloformyl group, a cyano group, an ester group including a lactone group, a benzyl, phenyl, tolyl, tosyl, sulfonyl group, an amino group (primary, secondary, tertiary), a sterol moiety, an isocyanate, a cyanate, a thioisocyanate, a thiocyanate, a carbamate, an azide, a diazo group, or a quinone group.
30. (New) The method of claim 14, wherein the substance is an organometallic compound, a β -hydroxy carboxylic acid, an inorganic acid or complex such as a metallocene, a nucleic acid.
31. (New) The method of claim 30, wherein the antibody is a polyclonal or monoclonal antibody, or a fragment thereof, a humanised or human antibody, an inhibitory or stimulatory antibody.

32. (New) The method of claim 14, wherein the substance is a protein or peptide.
33. (New) The method of claim 32, wherein the protein is a cytokine, a hormone, or an antibody.
34. (New) The method of claim 32, wherein the peptide is an oligopeptide comprising up to 20 amino acid residues
35. (New) The method of claim 34, wherein the oligopeptide is about 8, about 10 or about 12 amino acid residues in length.
36. (New) The method of claim 14, wherein the substance is a nucleic acid.
37. (New) The method of claim 36, wherein the nucleic acid is genomic DNA, cDNA, or mRNA, an oligonucleotide, or an oligoribonucleotide, wherein said nucleic acid encodes all or a fragment of a proteinaceous GTPase effector.
38. (New) The method of claim 37, wherein the encoding sequence is SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, or and 15.
39. (New) The method of claim 37, wherein the nucleic acid further comprises a gene therapy vector.

PATENT**IN THE UNITED STATES PATENT AND TRADEMARK OFFICE**

In re Application of:
Erik Nielsen, et al.

Serial No.: 10/088,549

Filed: March 15, 2002

For: ASSAY TO DETECT SUBSTANCES
USEFUL FOR THERAPY

Group Art Unit: Unknown

Examiner: Unknown

Atty. Dkt. No.: DEBE:007US

EXPRESS MAIL MAILING LABEL	
NUMBER	EL 503180194 US
DATE OF DEPOSIT	August 2, 2002

SECOND PRELIMINARY AMENDMENT**BOX PCT**

Commissioner for Patents
Washington, D.C. 20231

Sir:

Applicants respectfully submit this Preliminary Amendment in the above-referenced case. Consideration of this case in view of the amendments made herein is respectfully requested.

AMENDMENT

The Preliminary Amendment filed March 15, 2002 by the Applicants (copy enclosed) contained an error. An Amendment to the previously filed Preliminary Amendment is hereby requested.

On page 1 of the Specification, please delete the paragraph spanning lines 1 through 3 previously added by the Preliminary Amendment of March 15, 2002 and replace it with the following paragraph:

--This application claims priority to PCT/EP00/09130 filed on September 18, 2000, and EP 99 118 385.6, filed September 16, 1999. The entire content of both these applications are incorporated by reference.--


REMARKS

The specification has been amended to delete the incorrect priority data previously added by a Preliminary Amendment and to recite the correct priority data.

Should any fees under 37 C.F.R. §§ 1.16 to 1.21 be required, the Commissioner is hereby authorized to deduct said fees from Fulbright & Jaworski Deposit Account No. 50-1212/DEBE:007US.

The Examiner is invited to contact the undersigned attorney with any questions, comments or suggestions relating to the referenced patent application.

Respectfully submitted,

Respectfully submitted,

 REG No. 37,259 For

Steven L. Highlander
Reg. No. 37,642
Attorney for Applicants

FULBRIGHT & JAWORSKI L.L.P.
600 Congress Avenue, Suite 2400
Austin, Texas 78701
(512) 474-5201

Date: August 2, 2002

10/088549

Assay to Detect Substances Useful for the Therapy

The present invention relates to the use of effectors/regulators for Rab GTPases in *in vitro* and *in vivo* assays that recapitulate and measure the role of these effectors/regulators in membrane transport and membrane-cytoskeleton interactions in the endocytic pathway as novel targets to find therapeutic drugs to prevent or inhibit cancer cell growth and arrest cancer cell invasiveness as well as for stimulating and/or restoring endocytic transport and phagosome maturation in cells infected by intracellular parasites, which drugs are therefore useful for the therapy and optionally also the prophylaxis of 1) cancer and other proliferative (skin repair diseases such as psoriasis), invasive or cell migration disorders (endometriosis, atherosclerosis, inflammation and allergic diseases), 2) infectious (bacterial and viral) diseases, 3) diabetes, 4) Alzheimer's disease. In addition, the present invention is also directed to kits useful as a means to detect drugs suitable as anti-cancer and anti-infectious diseases drugs.

A class of molecules shown to play an important role in the regulation of intracellular transport and organelle function is represented by Rab proteins, small GTPases of the Ras superfamily, which are required in virtually every transport step which has been investigated. Similar to other GTPases, these molecules use the conformational change induced by GTP hydrolysis to regulate downstream events necessary for vesicle formation, docking and fusion. In the GTP-bound, active form Rab proteins bind to effector proteins and in this way transmit their signal to the transport machinery. For example, in membrane docking and fusion, Rab proteins regulate the activity of SNAREs. SNAREs are integral membrane proteins that by pairing on opposite membranes engaged in docking lead to membrane fusion. The pairing of SNAREs requires the activity of Rab proteins and therefore of Rab effectors. For example, endosome membrane docking requires the presence of the Rab5 effector EEA1 which, upon bridging the two opposite membranes, allows SNAREs to pair in trans, thus leading to membrane fusion (Chistoforidis et al., 1999a). The role of Rab proteins is not restricted to membrane docking and fusion but recent data provide evidence that Rab proteins also regulate the association with and motility of vesicles along cytoskeletal filaments. Membrane-cytoskeleton interactions play an important role in

determining the intracellular distribution and motility of organelles and transport vesicles. The cytoskeleton is made of different types of filaments, actin filaments and microtubules being the best characterised. Actin filaments and microtubules undergo continuous remodelling to serve the needs of the cell in cell architecture, motility, and organelle movement. Microtubules play a pivotal role in cell division but also constitute tracks along which cargo is transported across long distances such as between the periphery and the centre of the cell, from the basal to the apical pole in epithelial cells, or along the axon in neurons. Actin filaments are well known to function in muscle contraction. They are also important for cell motility, cell shape and vesicular transport. Different proteins are necessary to attach the organelles and transport vesicles to, and move them along, these filaments. Rab5 has recently been demonstrated to regulate the attachment of endosomes to, and motility along, microtubules. This suggests that Rab5, and in general members of the Rab proteins family, regulate various aspects of intracellular transport, including the ability of organelles to move along cytoskeletal tracks.

The identification of a large number of effectors and regulators for the small GTPase Rab5 provides a molecular explanation for the multiplicity of functions of Rab5 and allow to predict similar mechanisms for other Rab GTPases. Rab5 regulates a molecular network of several effector proteins, each contributing a specific function in membrane organisation, vesicle formation, vesicle and organelle movement, membrane docking and fusion. Most importantly, by functioning in a cooperative fashion Rab5 effectors modify the membrane environment and thus contribute to the biogenesis of the early endosome membrane. This mode of action is exemplified by the following mechanism elucidated by the present inventors. Upon activation by the effector/exchange factor Rabaptin-5/Rabex-5 complex, Rab5 locally recruits and activates phosphoinositide PI3-Kinases, leading to the generation of PI(3)P and consequently allowing the membrane recruitment of other Rab5 effectors (e.g. EEA1; see below) that bind to both Rab5:GTP and PI(3)P (Christoforidis et al., 1999b). Furthermore, Rab5 effectors are engaged in the formation of oligomeric complexes on the early endosome membrane (McBride et al., 1999). By the same criteria, other Rab proteins present in the early endosomes would be expected to recruit multiple effectors within a separate membrane environment. Consistent with this, studies using (green fluorescent protein) GFP-tagged Rab5, Rab4, and Rab11 have demonstrated that these GTPases are present in separate sub-compartments of the early endosome membrane. Endosomes are therefore organised as a mosaic of different Rab-domains created through the recruitment of specific effector proteins, which co-operatively act to generate a topologically restricted and functionally specialized environment on the endosome membrane.

Laying at the core of the organelle biogenesis and membrane transport mechanisms, Rab GTPases and their effectors are placed in a strategic position to regulate and mediate the trafficking of cellular membrane and soluble components along the biosynthetic and endocytic pathways. Their function includes the regulation of transport of cellular constituents (proteins and lipids) that are implicated in pathological alterations. Consequently, the present inventors' understanding of the mechanisms underlying membrane trafficking and membrane-cytoskeleton interactions, has applications in biotechnological research. Applications are in particular possible for those diseases where modulation of the membrane trafficking properties of endosomes provides an opportunity for therapeutic intervention. The main research areas for which this knowledge has proven to have important implications are: 1) cancer and other proliferative (skin repair diseases such as psoriasis), invasive or cell migration disorders (endometriosis, atherosclerosis, inflammation and allergic diseases), 2) infectious (bacterial and viral) diseases, 3) diabetes, 4) Alzheimer's disease. In the following section, the Rab GTPases and Rab effectors machinery will be presented in the context of the development of a therapeutic strategy for the treatment of these diseases.

1. Cancer

The molecular analysis of the transport machinery represented by Rab effectors operating in the endocytic pathway is relevant to two related aspects of the molecular mechanisms underlying tumor growth: intracellular signalling and cell spreading.

Intracellular signalling

Cell growth is regulated by growth factors that by binding specific receptors on the cell surface trigger a signalling cascade that culminates with the regulation of gene expression. Growth factors and their receptors are important mitogens both in normal and in transformed cells. Mutations in their genes or overexpression can cause tumors or stimulate tumor cell growth. For example, many tumors overexpress growth factors and surface tyrosine kinase receptors thus enhancing tumor cell growth. Growth factors and growth factor receptors have therefore been considered as excellent targets for new therapeutic drugs in the treatment of cancer and other disorders of excessive cellular proliferation. For example, the Herc2 receptor implicated in metastatic breast cancer provided the target for the development of Herceptin, a monoclonal antibody therapeutic from Genentech. The strategies pursued so far include the search for compounds that i) block ligand-receptor interactions, ii) the kinase activity of the receptors or the subsequent interactions with downstream factors of the signalling cascade. Another approach

consists of the search for drugs that modulate receptor trafficking through the endosomal system and result in receptor downregulation at the plasma membrane. Growth factor receptors and their bound ligands are internalised into endosomes, and endocytic trafficking of these molecules plays a critical role not only in attenuating the signalling response but also in establishing and controlling specific signalling pathways. The ability of growth factors to trigger signaling depends on the surface appearance of their receptors. This is determined by the kinetics and extent of receptor 1) endocytosis, 2) recycling to the cell surface and 3) transport to, and degradation in, late endocytic compartments (Sorkin et al., 1991). Molecules that can modulate these events and decrease the fraction of growth factor receptors on the plasma membrane are therefore valuable tools for an anti-cancer therapy. For example, enhanced degradation of epidermal growth factor receptor correlates with the lack of epidermal growth factor- induced proliferation and mitogen-activated protein kinase stimulation (Caraglia et al., 1999). Given their established role in the regulation of endosome structure and function, Rab GTPases and their network of Rab effectors represent an ideal target for screening of molecules that can reduce the surface content of growth factor receptors and, consequently, inhibit the growth properties of tumor cells. Rab5 for example regulates the internalisation of receptors from the plasma membrane into clathrin-coated vesicles (CCV) as well as the subsequent delivery of these carriers to early endosomes. Other Rab proteins such as Rab4 and Rab11 regulate the sorting function of early and recycling endosomes and, consequently, the recycling to the surface of growth factor receptors. The small GTPase Rab7 regulates transport of molecules from early to late endosomes and lysosomes, the degradative endocytic organelles.

The impact of Rab GTPases and their effectors/regulators on mitogenic signaling also includes the trafficking of signaling molecules downstream of growth factor receptors. An increasing number of components of the signalling machinery have recently been localised to early endocytic organelles. This remarkably specific localisation is very likely an important element in the signalling process. Such localisation depends on the structural and functional properties of the early endosomes, which in turn depends on the activity of Rab GTPases and their corresponding effectors/regulators. For example, the localisation of FYVE finger proteins depends on the synthesis of PI(3)P on the early endosome membrane (Christoforidis et al., 1999b). In the absence of this phosphoinositide these proteins are no longer capable of residing on early endosomes and are therefore released into the cytosol. Several novel signaling molecules possess a FYVE finger and, consequently, their ability to reside on early endosomes and to participate in the signaling cascade depends on the synthesis of PI(3)P. This concept is

corroborated by recent data indicating that hVPS34, the PI3-K that produces PI(3)P is required for mitogenic signaling (Siddhanta et al., 1998). The synthesis of PI(3)P is subjected to the regulation by Rab5, since PI-3 kinases (including hVPS34) are Rab5 effectors (Christoforidis et al., 1999b). Indeed, recent studies of the present inventors have demonstrated that the production of PI(3)P is stimulated by Rab5 and inhibited upon removal of Rab5 from endosomes. These observations therefore establish a direct link between the activity of Rab5 and its effectors such as PI-3 kinases and the membrane recruitment of signalling molecules to early endosomes and their ability to function in the signalling process.

Cell spreading

The growth of human tumors depends not only on the proliferative advantage characteristic of cancer cells but also on the ability of these to invade tissues and to colonise different organs. The ability of tumors to form metastatic tumours depends on the invasive properties of transformed cells. In addition, cancer cells require a higher supply of nutrients and, therefore, require de novo formation of blood vessels. Clearly, the spreading of endothelial cells during angiogenesis that occurs concomitantly with tumor growth also requires an increase in cell motility. Cell locomotion is therefore a central mechanism in tumor cell invasiveness and metastasis. On the one hand, cell spreading and invasiveness depend on the signalling function of various growth factor and growth factor receptors. In many different cell types, including tumor cells, EGFR signaling produces a pleiotropic response that includes mitogenesis or apoptosis, enhanced cell motility, protein secretion, and differentiation or dedifferentiation. The formation of new blood vessels is instead tightly regulated by specific growth factors that target receptor tyrosine kinases (RTKs) such as vascular endothelial growth factor (VEGF) (McMahon, 2000). A novel approach to treat human cancers emerging from these studies consists in the development of therapeutic strategies aimed at blocking tumor cell invasiveness and deregulated angiogenesis through the inhibition of various growth factor receptor signaling pathways (both in tumor cells and in the neighbouring tissue such as in endothelial cells). The potential use of Rab GTPases and their effectors/regulators as targets for the screening of molecules that can reduce the signaling function of growth factor receptors applies therefore not only for the mitogenic signaling but also for the spreading of tumor cells. On the other hand, the use of Rab GTPases and their effectors/regulators can also be extended to the cellular machinery that executes the signaling program resulting in increased cell locomotion. Cell locomotion occurs by a complex mechanism involving the coordinated activity of both the cytoskeleton - actin and microtubules - and membrane trafficking - especially endocytic and recycling structures. The structural/functional

properties of endosomes are important elements of the cell motility apparatus. For example, the endocytic cycle between endosomes and the plasma membrane sustains the deposition of adhesion molecules to the leading edge of motile cells. Consequently, the function of molecules that play an important role in endocytic transport can be rate-limiting important factors for the survival and spreading of cancer cells. This proposal is supported by recent studies that demonstrate that the small GTPase Rab5 is required for cell motility. The use of Rab GTPases and their effectors/regulators is therefore here applied for the screening of molecules that can reduce the ability of the cell to respond to mitogens through increased cell motility and, consequently, inhibit the spreading of tumor cells.

The described application of Rab effector intervention as a means of inhibiting or regulating cell motility defines the potential of the intervention for the treatment of other diseases that depend on cell locomotion such as endometriosis, atherosclerosis and monocyte migration into plaques, asthma and allergic diseases, and any other disease dependent on cell migration or locomotion.

2. Infectious diseases

Another area according to the present invention where membrane trafficking molecules, i.e., Rab GTPases and their effectors/regulators, can serve as therapeutic targets is that of infectious diseases. Cells internalise soluble ligands through endocytosis and large particles through phagocytosis. The latter mechanism is exploited by parasites that in this way gain access to the endomembrane system of the host cell. Increasing numbers of studies indicate that phagocytosis is regulated by the same molecular principles that govern endocytosis and recycling. For example, amphiphysin II and dynamin 2, which participate in receptor-mediated endocytosis, are also required for phagocytosis at the stage of membrane extension around the bound particles. Since the phagocytic process results in the internalisation of a large portion of the plasma membrane, mechanisms exist to ensure rapid renewal of plasma membrane. Given the role of Rab proteins in the regulation of membrane transport, it is obvious that they play an important role in the phagocytic cycle as well. Indeed, recent studies have shown that Rab11 operates in an endocytic compartment which is essential for phagocytosis. The function of endocytic Rab GTPases and their corresponding effectors/regulators are therefore expected to play an important regulatory role in the host cell invasion by parasites mediated by phagocytosis (Rab5) and in the compensatory membrane recycling (Rab4, Rab11) sustaining this process.

Once internalised, engulfed parasites or particles reside intracellularly in phagosomes. Under

normal conditions, phagosomes mature by docking and fusing progressively with early endosomes, late endosomes and lysosomes, thus causing the destruction of their content due to the low pH and increasing exposure to hydrolytic enzymes. However, it is well established that certain microbes can sabotage the cellular defence mechanisms and survive intracellularly. This is the case for example for *Mycobacterium tuberculosis* that once internalised by phagocytosis resides within the phagosome without coming in contact with harmful organelles such as lysosomes. A possible strategy to facilitate the killing of the pathogen is to re-activate the membrane trafficking route, thus resuming the maturation of the phagosome and, therefore, the exposure of the internalised bacterium to low pH and harmful hydrolytic enzymes. The strategy should focus on different Rab proteins each regulating a distinct stage of phagosome maturation (i.e. Rab5, Rab4, Rab11, Rab7).

Another area where Rab GTPases and their effectors can be used for therapeutic intervention is that of viral infection, especially HIV. AIDS continues to be cause of many deaths both in developed and under-developed countries. It has been recently established that, besides CD4, members of the family of chemokine receptors are required as co-receptors, which in conjunction with CD4, allow the virus to enter cells. Downregulation of chemokine receptors by endocytosis and inhibition of receptor recycling protects cells from HIV. These observations imply that receptor internalization and inhibition of receptor recycling represent novel therapeutic strategies to prevent HIV infection and therefore combat AIDS. Since Rab GTPases and their effectors/regulators regulate the trafficking of receptors to the cell surface, these molecules represent new targets for therapeutic agents aimed at decreasing the surface expression of HIV receptors and therefore inhibit HIV entry into cells and prevent HIV infection.

3. Diabetes

Another area where the endocytic trafficking apparatus has potential commercial applications is the trafficking of glucose transporters and the defect in this process, which is a cause of diabetes. In fat and muscle cells, stimulation by insulin causes the relocation of glucose transporters (e.g. GLUT4) from intracellular stores to the cell surface. These intracellular stores are thought to be specialised membrane domains of the recycling endomembrane system. Insulin sensitivity depends on the surface levels of such glucose transporters (GLUTs). Under physiological conditions, the increase in muscle insulin sensitivity of glucose transport observed after exercise is due to translocation of more GLUT-4 to the cell surface. Under pathological conditions, GLUTs trafficking is perturbed. Certain forms of diabetes are in fact caused by deficiency in the

surface appearance of the GLUTs. This implies that the elucidation of the transport machinery that regulates the trafficking of membrane proteins, including GLUTs, along the endocytic and recycling pathway offers therefore interesting opportunities for intervention in diabetes. In particular, the discovery of Rab effectors that play a role in the transport to endosomes and from endosomes to other destinations, e.g., the cell surface or to degradative compartments, identifies potential targets for the screening of drugs that, by altering the trafficking properties of GLUTs, can restore or stimulate their delivery to the plasma membrane in response to insulin.

4. Alzheimer's Disease

At the core of the molecular mechanism underlying the generation of Alzheimer's disease lies the aberrant processing of the Amyloid Precursor Protein (APP) by β -secretase to form $A\beta(1-42)$. It is at present unclear where precisely the aberrant cleavage occurs, but endosomes have been implicated as the compartment or one of the compartments where processing of APP occurs. This implies that 1) APP is transported to endosomes and follows the endocytic/recycling pathway and 2) β -secretase activity is localised to endosomes. Since Rab GTPases and their effectors/regulators regulate the trafficking of molecules (lipids and proteins) in the endocytic pathway, these molecules represent new targets for therapeutic agents aimed at decreasing the presence of APP in endocytic compartments and/or the presence of β -secretase activity in endocytic compartments, therefore inhibiting the generation of beta-amyloid and retarding and or preventing the onset/progression of Alzheimer's disease.

Based on the above experimental results, the object of the present invention was to develop an assay useful for the detection and identification of molecules (pharmaceutical drugs). In order to achieve this object, the present inventors developed the strategy that molecules (proteins) interacting with GTPases such as GTPases of the Rab family and, thus, functioning at the level of endosome trafficking and coordinating the interaction of these organelles with the cytoskeleton, can provide novel targets for molecules (pharmaceutical drugs) aimed at inhibiting cell locomotion and invasiveness (and thereby cancer) as well as infectious (both viral such as AIDS and bacterial and also eukaryotic), allergic, and inflammatory diseases, diabetes, Alzheimer's disease, endometriosis, and atherosclerosis. A further object of the present invention was to provide a kit comprising molecules interacting with GTPases (effector or regulator proteins/molecules; for exact definitions see further below), which kit is useful as a means for the detection and identification of pharmaceutical drugs exhibiting properties making them preferred

drugs for treating cancer, inflammatory, allergic, and infectious diseases (eukaryotic, prokaryotic and viral) including AIDS but also atherosclerosis, endometriosis, diabetes and Alzheimer's disease. Hereinafter, these diseases are termed "the above mentioned diseases".

5 The following definitions are given in order to more fully specify the terms as used in the present application. **"Invasiveness"** refers to the migration and spreading of cancer cells or other cells such as endothelial cells (rather than the invasion of bacteria). **"GTPase-interacting protein"** or **"regulator/regulatory protein/regulatory molecule"** refers to any protein that interacts with the GTPase under any nucleotide conformation and for any purpose. GTPase-interacting
10 proteins/regulators regulate the nucleotide-bound state of the GTPase, i.e., the GTPase activating protein (GAP) interacts with the GTP-bound GTPase and down regulates the activity of the GTPase by stimulating the hydrolysis of GTP. A GDP/GTP exchange factor promotes nucleotide exchange, thereby activating the GTPase. **"Effector"** or **"effector/effector protein/effector molecule"** is defined to mean a protein or a protein complex that interacts with the GTPase only
15 if the GTPase is in the GTP-bound, also called active, form, that by binding stabilises this conformation and that mediates the "effect" of that GTPase. Effectors transmit the function of the GTPase, Rab is upstream and recruits the effectors which do the work. Regulators instead switch the Rab on and off. Rab-on (=GTP) binds the effectors and is functional. Thus, both effectors and regulators may therefore also be termed GTPase effectors and GTPase regulators,
20 respectively. Additionally, it is apparent for the skilled reader that the term **"regulator"** is broader than the term **"effector"** and includes the latter.

The above mentioned strategy implies the first step of employing known effectors/regulators or identifying novel effectors/regulators of Rab or of other (small) GTPases, specifically regulating
25 endosome trafficking and endosome-cytoskeleton interactions, and the additional step of designing an *in vitro* and/or *in vivo* assay to screen for inhibitors or activators of these effector/regulator molecules. In particular, the strategy includes the identification of substances useful as pharmaceutical drugs due to their interference with the process of cell locomotion, i.e., it includes the identification of inhibitors of membrane-cytoskeleton interactions.

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The above object in mind, the present inventors established an assay system based on the use of such effectors/regulators, as defined hereinbefore, as targets for pharmaceutical drugs useful in the therapy of cancer and infectious diseases and any other disease which affects or requires the function of the endocytic/recycling trafficking machinery. According to a preferred embodiment

of this aspect of the present invention, an assay is provided using both an effector/regulator and a Rab protein (or any other small GTPase). Accordingly, one aspect of the present invention is the use of the above mentioned GTPase effector/regulator molecules or proteins as targets in an *in vitro* or *in vivo* assay system to detect substances useful as a pharmaceutical/therapeutic agent for the prophylaxis and the treatment of the above mentioned diseases (including cancer, allergic, inflammatory, and infectious diseases, diabetes, endometriosis, atherosclerosis and Alzheimer's disease). Another object of the present invention is a kit useful for carrying out the assay of the invention. The kit necessarily contains at least one type of effector/regulator molecule. At least one GTPase effector/regulator molecule is either native and biochemically purified from a vertebrate, or recombinant and biochemically purified from bacterial cultures, from yeast cultures, or from other cultured eukaryotic cells, in either case labelled by a covalent modification or radioactivity suitable for use in the assay.

According to a preferred embodiment of this aspect of the present invention the kit additionally contains at least one type of GTPase and/or endosomal membrane fractions obtained by subcellular fractionation and labelled by internalisation of transferrin, either fluorescently labelled or labelled by any other modification that allows its detection, and/or cytosolic extracts, and/or an ATP-regenerating system and/or a number of chemicals to be tested for their suitability as a drug against any of the above mentioned diseases (cancer, endometriosis, diabetes, atherosclerosis, allergic, infectious, inflammatory diseases). Other specific and preferred embodiments of both the kit and the use according to the present invention may be readily taken from the claims attached hereto.

Drugs exhibiting any of the above activities include those molecules (including peptides) mimicking the peptide structure of the respective target molecule. Typical representatives of the above classes are molecules/substances that carry one or more of the following functional groups: a halide atom bound to an alkyl, alkenyl, alkynyl, or aryl residue, an alcohol group (primary, secondary, tertiary), an ether group, a carbonyl function (aldehyde or ketone), a carboxylic acid group, a carboxylic anhydride group, a carbamoyl group, a haloformyl group, a cyano group, an ester group including a lactone group, a benzyl, phenyl, tolyl, tosyl, sulfonyl group, an amino group (primary, secondary, tertiary), an isocyanate, a cyanate, a thioisocyanate, a thiocyanate, a carbamate, an azide, a diazo group, and a quinone group. Other representatives of suitable drugs are organometallic compounds, sterol moiety(ies)-containing molecules, β -hydroxy carboxylic acids, inorganic acids, and complexes such as metallocenes; nucleic acids,

cytokines, hormones, antibodies, or oligopeptides comprising up to 20, preferably 8, 10, or 12, amino acid residues.

Preferably, the antibodies are polyclonal or monoclonal antibodies, or fragments thereof, humanised or even human antibodies (obtained for example by a phage display method), inhibitory or stimulatory, raised against and targeted towards any of the aforementioned GTPase effectors. A preferred nucleic acid is genomic DNA, cDNA, or mRNA, or a fragment thereof, an oligonucleotide, an oligoribonucleotide, all being based on or derived from any of the GTPase effector gene sequences having any of the sequences as depicted in SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, and 15, or gene therapy vectors derived from the aforementioned GTPase effector gene sequences.

The advantage (and the gist) of the present invention consists in (1) the development of a comprehensive screening method based on molecular networks, (2) the sensitivity of the assays, and (3) the selectivity of the targets. The activity of the factors under control of the central switch can be responsible for coordinating various interconnected functions such as (a) endocytosis, (b) the distribution, i.e., sorting of proteins and lipids within endosomes and from endosomes to other intracellular locations (e.g. interactions of phagosomes with endosomes and lysosomes), (c) the motility of endosomes along the cytoskeleton, and (d) cell migration. This multiplicity of factors responsible for a wide number of cellular functions is defined as a *molecular network*. The present invention resides in enabling the skilled person to study the molecular network of many different GTPases, in particular those GTPases localised to endosomes, e.g., Rab5, Rab4, Rab11, Rab7, Rab17, Rab18, Rab22, as a whole in order to establish various (GTPase) effector/regulator proteins that are suitable as targets for pharmaceutical drugs. In this approach, the GTPase itself will not serve as a target molecule because its activity is so crucial for cell homeostasis that it would likely invariably result in toxic effects. Rather, the effector/regulator molecules specifically functioning in protein and lipid sorting, endosome motility and endosome-cytoskeleton interactions will be the target. For example, compounds that affect cell motility could be screened more specifically without affecting other important cellular functions (i.e. endocytosis). In the alternative, and according to a preferred embodiment of the present invention, the target may well be one of the effectors/regulators as defined above, but additionally a GTPase such as Rab5, Rab4, Rab7, Rab11, or Rab17, will be present in the assay system.

The present inventors have functionally characterised several Rab GTPases (e.g., Rab5, Rab4, Rab11, Rab 7, Rab17, Rab18, Rab20) and effectors/regulators of these GTPases, localised to the early endosomes and the recycling endosomes, which GTPases and effectors/regulators, respectively, are thus suitable for the assay according to the invention. These Rab proteins sequentially control transport in the endocytic and recycling pathway. Rab5 controls transport from the plasma membrane to early endosomes whereas Rab4 and Rab11 function in protein sorting and recycling from early and recycling endosomes back to the plasma membrane. The expression of Rab17 is restricted to polarised cells such as epithelial cells and neurons. This latter GTPase regulates trafficking through the apical recycling endosome, thus contributing to the generation and maintenance of cell polarity. Thus, effectors/regulators of all these GTPases and the respective GTPases (in combination with their effectors/regulators) may be employed in accordance with the present invention in an *in vitro* or *in vivo* assay system as a target in order to screen for substances that are useful as a pharmaceutical agent for the prophylaxis and treatment of the above mentioned diseases, i.e., cancer (benign tumours, malignant tumours, carcinomas, sarcomas, melanomas, leukemias, gliomas, or neuroblastomas), diabetes, Alzheimer's disease, endometriosis, atherosclerosis, allergic, inflammatory, and infectious diseases such as AIDS, tuberculosis, cholera, malaria, pseudotuberculosis, gastroenteritis, enteric fever, typhus, and any other infectious diseases caused by an infectious agent that infects cells by the endocytic route and resides intracellularly in phagosomes escaping the cellular killing mechanisms (in this regard, it should be recalled that the GTPases per se and alone can not serve as a target for the drugs to be detected; rather, the target for the drug is an effector/regulator molecule). Other examples of infectious diseases are those diseases caused by the following pathogens (bacteria or organisms): Mycobacterium, Staphylococcus, Toxoplasma, Trypanosoma, Listeria, Salmonella, Legionella, Leishmania, Coxiella, Shigella, Yersinia, Neisseria, Vibrio, Bartonella, and any other intracellular pathogen that resides in intracellular phagosomes and escapes cellular killing mechanisms.

As the proteins described herein, i.e., the Rab proteins and their effectors/regulators, are, unless exceptions, ubiquitously expressed, pharmaceutical agents for the prophylaxis and treatment of tumours of any cellular origin can be screened for and detected according to the present invention. Specifically, the following tumours may be mentioned: lung carcinoma, osteosarcoma, lymphoma, leukemia, soft tissue sarcoma, breast carcinoma, bile cancer, cervix carcinoma, cancer of the (small) intestine, of the kidneys, of the cavity of the mouth, penis carcinoma, ovary cancer, stomach cancer, cancer of the tongue, brain cancer, bladder carcinoma, prostate

carcinoma, liver carcinoma, and carcinoma of the pancreas and every tumor that invades other tissues and organs.

Drugs with anti-cancer and/or an activity against any of the other above mentioned diseases include those molecules (including peptides) mimicking the peptide structure of the respective target molecule. Typical representatives of the above classes are molecules/substances that carry one or more of the following functional groups: a halide atom bound to an alkyl, alkenyl, alkynyl or aryl residue, an alcohol group (primary, secondary, tertiary), an ether group, a carbonyl function (aldehyde or ketone), a carboxylic acid group, a carboxylic anhydride group, a carbamoyl group, a haloformyl group, a cyano group, an ester group including a lactone group, a benzyl, phenyl, tolyl, tosyl, sulfonyl group, an amino group (primary, secondary, tertiary), an isocyanate, a cyanate, a thioisocyanate, a thiocyanate, a carbamate, an azide, a diazo group, and a quinone group. Other representatives of suitable drugs are organometallic compounds, sterol moiety(ies)-containing molecules, β -hydroxy carboxylic acids, inorganic acids, and complexes such as metallocenes; nucleic acids, cytokines, hormones, antibodies, or oligopeptides comprising up to 20, preferably 8, 10, or 12, amino acid residues. Effectors transmit the function of the GTPase, Rab is upstream and recruits the effectors which do the work. Regulators instead switch the Rab on and off. Rab-on (=GTP) binds the effectors and is functional.

To elucidate the molecular mechanism whereby these GTPases exert their function in membrane trafficking and cytoskeleton organisation, the present inventors have developed a method to biochemically identify Rab5 protein effector molecules (Chistoforidis et al., 1999a). This method is based on previously described affinity chromatography (Amano et al., 1996) but has been optimised to result in the specific elution and large-scale purification of Rab effectors and GTPase interacting proteins in general, in amounts sufficient for both their identification by microsequencing techniques and their functional characterisation (Chistoforidis et al., 1999a).

In addition to that method, the present inventors have developed a novel method. This method is described in Example 3 below. With this method, the inventors have re-purified several of the proteins that bind Rab5. Using this modified affinity chromatography procedure, more than 20 effector and/or regulator proteins interacting directly or indirectly with Rab5 (Figure 1a in (Chistoforidis et al., 1999a)) have been identified and demonstrated to exhibit functional activity as shown by their ability to substitute cytosol in an *in vitro* endosome fusion reaction (Chistoforidis et al., 1999a). Some of these molecules such as Rabex-5 regulate the activity of

Rab5 (i.e., its nucleotide state) (Horiuchi et al., 1997). Others such as Rabaptin-5 and EEA1 participate in the endosome docking and fusion process (Stenmark et al., 1995). Still others regulate the ability of endosomes to move along microtubules, and all of these proteins are suitable candidates as a target for the detection of chemical drugs (compounds) that may turn out to be highly efficient agents to combat the above mentioned diseases.

In addition, expression of an activated mutant of Rab5 (Rab5Q79L) induces an increased recruitment of actin on the expanded endosomes suggesting that Rab5 either directly or indirectly regulates the attachment/nucleation of actin filaments with the endosome membrane. Based on the above data, in particular in view of this large number of Rab5 interacting molecules (more than 20), the present inventors concluded that the activity of Rab5 is not restricted to endocytic membrane docking and fusion. Most importantly, they also concluded that modulating the activity of Rab5, and therefore of its effectors/regulators, would result in changes in organelle motility *in vivo* and in cell motility. This is perfectly consistent with the effect observed by the present inventors that expression of activated Rab5 leads to an increase in motility of fibroblasts and endothelial cells, implicating this GTPase in the regulation of cell migration. The effector/regulator protein(s) responsible for this activity (to regulate membrane trafficking and membrane-cytoskeleton interactions including the ability of endosomes to move along microtubules) are therefore, in accordance with the present invention, specific targets for drugs aimed at inhibiting the ability of the Rab5 machinery to sustain cell migration.

Similar experiments have been and are being conducted on two other GTPases functioning in endosome trafficking, Rab4 and Rab11. Several candidate proteins for Rab4 effectors/regulators have been purified and one of these proteins has been sequenced and identified as the human homologue of the yeast Adenyl Cyclase-associated protein CAP (Field et al., 1990). This protein is a component of the Ras-activated cyclase complex in yeast and therefore is required for the proliferative signal in yeast. In addition, this protein has been implicated in the interaction with the actin cytoskeleton, since the carboxy terminus of CAP binds actin. Another protein which has been identified is Rabenosyn-5, a Rab5 effector. The realisation that this molecule binds both Rab5 and Rab4 but does not interact with Rab11, suggests that it may control trafficking between the Rab5 and Rab4 sub-compartments within early endosomes, and therefore regulate the sorting of proteins trafficking through this organelle. This hypothesis is consistent with the new experimental data recently obtained (see below).

Given the role of Rab proteins in the regulation of membrane transport and membrane-cytoskeleton interactions, the effectors/regulators for these small GTPases are targets for pharmaceutical drugs aimed at (1) inhibiting cell invasiveness and (2) enhancing cell-defence mechanisms against intracellular pathogens, such drugs therefore being valuable weapons in the battle against tumor (and cancer) and all other of the above mentioned diseases. For example, it may be possible to inhibit the stimulatory activity of Rab5 to reduce cell motility and consequently to reduce metastasis and the progression of cancer.

A strategy similar to that established and described above for Rab5 has been applied for other Rab proteins regulating endocytic trafficking. For example, the small GTPases Rab4, Rab11, and Rab17 reside onto and regulate trafficking through the early and recycling endosomes. Given that in motile cells recycling endosomes preferentially deliver membranes to the leading edge of the cell and given that the deposition of adhesion molecules to the leading edge of motile cells depends on the endocytic cycle between endosomes and the plasma membrane, the established functional role of Rab proteins functioning on endosomes, including Rab4 and Rab11 characterised by the present inventors, implies that they may participate in the regulation of cell motility. Therefore, the effectors/regulators of these and other endosomal GTPases will also represent novel potential targets according to the instant invention.

On the other hand, further Rab effectors/regulators to be identified and characterised in the future will also act in various aspects of endosome function, including endosome and cell motility, and endosome-phagosome interactions. An approach to identify further effectors/regulators has been described (Chistoforidis et al., 1999a), and the procedure described by the authors is similarly suitable to detect, identify, and characterise further, as yet unknown, effector/regulator molecules (proteins) of small GTPases of the Rab family.

The multiplicity of Rab5 (or any other Rab GTPases) effector/regulator molecules serves to execute distinct functions in endosome dynamics and cell dynamics all coordinated by the GTPase switch of Rab5 (or any other of the GTPases). It is therefore a "package" also referred above as a network of molecules that is necessary to coordinate different functions. These molecules can operationally be divided into the following classes on the basis of either their established identity and/or function and on the basis of their possible function for which different claims in drug discovery can be proposed. The effectors/regulators of all classes may be used for

the assay as defined herein, i.e., for the detection of suitable drugs for the treatment of any of the above mentioned diseases.

Class 1: Representative effectors/regulators: Rabaptin-5, Rabaptin-5b, Rabex-5, RN Tre, EEA1, PI 3-K, Inositol Polyphosphate 4-phosphatase and Inositol Polyphosphate 5-phosphatase.

Function: endocytic trafficking

Assay: endosome fusion, endosome motility

This class of Rab5 effectors/regulators includes molecules that 1) have either been already described as functionally linked with Rab5 (Rabaptin-5, Rabaptin-5b, Rabex-5, EEA1, PI 3-K), or 2) have previously been identified and whose DNA and deduced protein sequence have been disclosed (RN Tre, Inositol Polyphosphate 4-phosphatase and Inositol Polyphosphate 5-phosphatase) but that have been now functionally linked to Rab5 by the present inventors. Rab5 effectors/regulators of this class 1 belong to the “package” also referred above as a network of molecules that is necessary to coordinate different functions of Rab5.

Since Rab5 regulates endocytosis, effectors/regulators of this molecule also control endocytosis as well as all functions controlled by Rab5; i.e., endocytic trafficking, intracellular distribution of endosomes, motility of endosomes along microtubules, and cellular motility. Consequently, class 1 Rab5 effectors should be considered potential targets for the therapeutic strategies above described in conjunction with the other classes of Rab5 effectors/regulators.

Class 1 includes

1) Regulators that play a role in the nucleotide cycle of Rab5. Rabex-5 has been identified as a nucleotide exchange factor for Rab5 (Horiuchi et al., 1997), and therefore converts the GTPase from the inactive, GDP-bound, to the active, GTP-bound form. Consequent to the activity of Rabex-5 is the interaction between Rab5 and the Rab5 effectors that transmit Rab5 function. The present inventors have discovered that RN Tre (Matoskova et al., 1996), a protein related to the *Tre* oncogene, is a Rab5 GAP, which discovery provides an interesting link between the signal transduction and membrane trafficking machinery. Most importantly, like Rabex-5 that regulates activation of Rab5, RN Tre regulates its inactivation and therefore represents an ideal target for drugs aimed at interfering (positively and negatively) with Rab5 function.

2) Effectors that play a role in endosome docking and fusion: Rabaptin-5 (Stenmark et al., 1995), Rabaptin-5b (Gournier et al., 1998), and EEA1 (Christoforidis et al., 1999a; Simonsen et al., 1998), or in the regulation of this process via the generation of phosphoinositide-3-phosphate (PI 3-K) which is necessary for the membrane recruitment of EEA1 (Christoforidis et al., 1999b).

5 Two distinct phosphatidyl-inositol 3-kinases (PI 3-K), hVPS34 and p85a/p110b, have been identified among the Rab5 effectors. The two kinases are differentially distributed along the endocytic pathway, and the activity of hVPS34 is specifically required for EEA1 membrane recruitment and endosome fusion. It has also been demonstrated that VPS34 regulates the motility of endosomes along microtubules, suggesting that PI 3-K activity is necessary to coordinate the function of various Rab5 effectors/regulators, including those controlling membrane-cytoskeleton interactions (Nielsen et al., 1999). PI 3-K are therefore considered to represent a particularly suitable target of drugs that block the motility activity. Similarly, targets other than PI 3-K may be identified among the Rab5, Rab4, and Rab11 effectors/regulators that couple the activity in membrane docking and fusion with the organelle dynamics. One such target coupling the activity in membrane docking and fusion with the organelle dynamics is Rabaptin-5 due to its capacity to couple Rab4 to Rab5 function.

The present inventors have determined that the list of Rab5 effectors includes two more proteins that play a role in phosphoinositide metabolism: Inositol Polyphosphate 4-phosphatase and Inositol Polyphosphate 5-phosphatase. First tests conducted by the present inventors have proven that both these phosphatases bind to a Rab5 affinity column and directly interact with Rab5 in a yeast two-hybrid system assay. The function of Inositol Polyphosphate 4-phosphatase and Inositol Polyphosphate 5-phosphatase is to catalyse the removal of the phosphate from the 4- and 5-position of the inositol ring, respectively. In the context of the Rab5 network of effectors/regulators, these proteins are functionally linked to the PI3-Ks, especially p85a/p110b, since Inositol Polyphosphate 4-phosphatase and Inositol Polyphosphate 5-phosphatase would use as substrate the product of this kinase. The discovery that 1) Rab5 regulates a sophisticated cycle of phosphorylation/dephosphorylation of phosphoinositides and 2) that phosphoinositides regulate the recruitment of factors playing a role in the endosome transport machinery as well as signaling molecules, implies that Rab5 and its effectors/regulators can be used as drug targets for diseases where alterations in the function of endosomes can result in therapeutic intervention.

The class 1 effector/regulator molecules may be used according to the present invention as drug targets in cancer on the basis of regulating cell motility and invasiveness. The inventors of the present invention have found out that Rab5 regulates the motility of endothelial cells in vitro (this result was the outcome of experiments described in the Examples below). Since the

Rabaptin-5/Rabex-5 complexes regulate the nucleotide cycle of Rab5, and since the kinases PI 3-K regulate the recruitment of effectors/regulators, these molecules have been identified as targets for drugs aimed at inhibiting the activity of Rab5 in cell motility. The class 1 effector/regulator molecules may also be used as drug targets in cancer on the basis of the signalling molecules regulating cell growth. Growth factors and growth factor receptors are transported through the endosomes before being degraded in the late endosomes and lysosomes. Molecules that regulate the trafficking of receptors along the endocytic pathway can be used as targets for drugs aimed at interfering with transport (i.e., clearance from the plasma membrane, inhibition of recycling, transport to endosomes where signalling molecules are located, increase transport to degradative compartments where receptor-ligand complexes are degraded) and, consequently, inhibiting mitogenic signalling. Finally, the class 1 effector/regulator molecules may also be used as drug targets in infectious diseases (see the above list exemplifying infectious diseases particularly suitable according to the present invention). The small GTPase Rab5 has been observed to accumulate on *Mycobacterium tuberculosis* phagosomes. Interestingly, the Rab5 effector Rabaptin-5 is not present on the phagosome membrane. Thus, it must be concluded that there are differences between endosomes and phagosomes in the utilisation of the Rab5 effectors/regulators. In the case of *Listeria monocytogenes* phagosomes, it has been observed that an upregulation in expression of the small GTPase Rab5 occurs in the course of infection of macrophages by a *Listeria monocytogenes* non-lytic mutant. This increased Rab5 recruitment results in an increased fusogenicity of the phagosomes with endosomes leading to intracellular killing of the bacterium. Accordingly, in order to detect and identify substances suitable for the therapeutic treatment of pathogen infections, the Rab5 effector/regulator responsible for this effect may be used as a target in the assay described in detail further below. In this case, the upregulation of the host cell defence mechanism could be accomplished by a suitable drug. The activator of Rab5, Rabex-5, that functions as a nucleotide exchange factor and converts the protein into the GTP-bound active form (Horiuchi et al., 1997) is a particularly suitable drug target in this regard. By enhancing its activity (by means of a drug activating Rabex-5), the inventors have been able to up-regulate the Rab5 machinery, stimulate endosome fusion and facilitate phagosome-endosome interactions. Alternatively, other Rab5 effectors/regulators (still to be identified) playing a role in endosome trafficking could serve as targets.

Class 1 as well as the other classes of Rab5 effectors described below may include molecules playing a role in endosome motility. The present inventors have found that Rab5 regulates the motility of early endosomes *in vivo* and both the attachment of early endosomes to microtubules

and the motility of early endosomes along microtubules *in vitro* (Nielsen et al., 1999). New data is that a Rab5 eluate is sufficient to reconstitute motility in an endosome-microtubule *in vitro* motility assay and in the absence of cytosol. Therefore a Rab5 effector(s) is/are sufficient to regulate or switch on a microtubule motor, or a Rab5 effector(s) is/are a microtubule motor.

5 For this function we make three statements to support a claim:

1. Rab5 is required to bind certain or possibly all early endosomes to microtubules.
2. Rab5 effectors and possibly their binding partners (i.e. a Rab eluate) are required for endosome motility on microtubules.
3. RabGDI blocks all endosome motility *in vivo* (microinjection of RabGDI).

10 These data demonstrate an essential role for Rab5 effectors in binding endosomes to microtubules and their subsequent motility along microtubules. Rab5 effectors can therefore be used as target molecules for drugs aiming to reduce endosome motility to block endosome cycling and, consequently, recycling of surface molecules necessary for cell motility.

15 Class 2

Representative effectors/regulators: Rabankyrin-5 and Rabenosyn-5 (p110 FYVE)

Function: endocytic trafficking, possible interaction with the actin cytoskeleton .

Assay: endosome fusion, endosome motility

20 This class includes at least two proteins which share the same PI(3)P-binding domain, the FYVE-finger. One protein is the human homologue of mouse Ankhzn (Ito et al., 1999), to be renamed Rabankyrin-5, given its ability to bind Rab5. The present inventors also found a Rabankyrin-5 splice variant (SV). Rabankyrin-5 contains ankyrin repeats. It is localised to early endosomes, binds Rab5, plays a role in endosome fusion. and is required for early endosome
25 fusion. As regards the binding to Rab-5, it is worth noting that the present inventors determined the Rab5 binding region to reside in the ankyrin repeats in the C-terminus of the protein (between residues 542 and 1075).

The sequence of the gene coding for human Rabankyrin-5 is depicted in the sequence listing
30 attached to this application (SEQ ID NO: 1) whereas the sequence of the protein itself is given in the sequence listing as SEQ ID NO: 2. The nucleotide and amino acid sequences of the SV gene and protein are indicated in the sequence listing as SEQ ID NO: 3 and SEQ ID NO: 4, respectively.

To the best knowledge of the present inventors, there is no prior art that would suggest a role for the gene/gene product of ankhzn - or of its human analogues - as a Rab5 effector with a role in endosome fusion and membrane trafficking. The second molecule is Rabenosyn-5, previously named p110FYVE. It is localised to early endosomes as well, binds Rab5 as well, and is also known to be complexed with and bind to the mammalian homologue of yeast Vps45p, a protein involved in transport to the vacuole. This binding demonstrates that Rabenosyn-5 has a role in SNARE-mediated membrane fusion. Rabenosyn-5 (p110FYVE) probably also plays a role in the regulation of transport and participates in the movement of endosomes along the actin and microtubule cytoskeleton.

The nucleotide and amino acid sequences of p110FYVE are given in SEQ ID NO: 5 and SEQ ID NO: 6, respectively. Further properties of the effector Rabenosyn-5 (p110 FYVE) are that it is essential in endosome fusion, fusion of clathrin coated vesicles derived from the plasma membrane with early endosomes. Over-expression of Rabenosyn-5 inhibits the processing of lysosomal enzymes.

The present inventors have found that Rabenosyn-5 is a Rab4 effector. It has been known that three small GTPases, i.e., Rab5, Rab4, and Rab11, exist in separate domains on the same endosome. The present inventors have been able to demonstrate that over-expression of Rabenosyn-5 brings the Rab5 and Rab4 domains together, without changing the distribution of Rab11, and changes the sorting properties of the endosome with respect to the cycling of certain receptors (e.g. transferrin receptor). This proves that Rabenosyn-5 has a function in receptor sorting and provides validation that manipulating Rabenosyn-5 has the potential to control receptor amount, distribution, and the kinetics of receptor cycling.

Class 3

Representative effectors/regulators: multi-protein complex including p100, p95, p60, p45, p25

Function: endocytic trafficking, possibly in polarised cells such as epithelial cells and neurons

Assay: endosome fusion, endosome motility

This complex consisting of the above 5 proteins (p100, p95, p60, p45, p25) and exhibiting the molecular weights of 100 kDa, 95 kDa, 60 kDa, 45 kDa, and 25 kDa, respectively, has been isolated by the present inventors. At least one of these molecules, or the entire complex, interacts either directly with Rab5 or indirectly through another Rab5-binding protein. The present

inventors have also determined that the complex is detected in a clathrin coated vesicles-enriched fraction. This subcellular fraction includes the vesicles that transport protein receptors from the plasma membrane to the early endosomes.

- 5 The structure (sequence) and function of the 5 protein components have been investigated to some extent and are more or less known now and form part of the present invention.

In detail, the C-terminal half of the 100 kDa component shares sequence homology (52%) with the conserved catalytic domain of GTPase activating proteins and spindle assembly checkpoint
10 proteins from yeast to mammals, while the N-terminal half shares homology (57%) with the conserved domain of serine/threonine protein kinases. Fusion between early endosomes is strongly inhibited when cytosol is immuno-depleted using anti-45 kDa component antibodies.

Immunofluorescence data show that the 45 kDa component is localised in intracellular structures
15 which are not EEA1 positive. Upon over-expression of Rab5Q79L though, there is partial co-localisation of these proteins on early endosomes.

The nucleotide and amino acid sequences of these proteins have been determined and are depicted in the attached sequence listing (p60: SEQ ID NO: 7 and SEQ ID NO: 8; p45: SEQ ID
20 NO: 9 and SEQ ID NO: 10; p25: SEQ ID NO: 11 and SEQ ID NO: 12, p100: SEQ ID NO: 13 and SEQ ID NO: 14; p95: SEQ ID NO: 15 and SEQ ID NO: 16, respectively). The p95 sequences are only partial sequences, however.

Examples

25 Example 1:

The following example describes the materials and methods used to measure the effect of a GTPase or mutant GTPase on the regulation of cell motility.

Expression constructs

30 Myc-rhoDV26G (Murphy et al., 1996) was cloned into the HindIII-XbaI sites of expression vector pHSVPUC (Geller et al., 1993) to generate construct: pHSV-myc-rhoDV26G. Myc-rhoDV26G cDNA was cloned in frame into the EcoRI-BamHI sites of expression vector pEGFP-C2 (Clontech) to generate a green fluorescent protein (GFP) fusion: GFP-myc-RhodV26G.

Transient transfection of rhoDV26G and human transferrin receptor

BHK-21 cells were trypsinized 24h prior to transfection and were seeded onto 11mm glass coverslips. Cells were infected with T7 RNA polymerase recombinant vaccinia virus (Bucci et al., 1992) and transfected with either T7-myc-rhoDV26G and T7-human transferrin receptor (T7-hTR) or T7-hTR alone (Murphy et al., 1996) using DOTAP (Boehringer Mannheim, Mannheim, Germany). The cell number and also the amount of DNA and lipid were constant. The final concentration of DNA per coverslip was 1µg.

Following transfection, the cells were incubated at 37°C, 5% CO₂, for 2h, cyclohexamide was then added for 1h. Cells were washed 8 times for 1min in preheated medium and rhodamine transferrin (50 µg/ml) was uptaken in a preheated humidified chamber at 37°C, 5% CO₂, for 20min. Coverslips were washed in preheated medium and then mounted into chambers for video microscopy (Bradke and Dotti, 1997; Bradke and Dotti, 1996) as described below. TLVM (time lapse video microscopy) was carried out immediately. Hydroxyurea was present at all times to prevent late viral gene expression.

Transient transfection of T7-rhoDV26G and T7-gfp

To monitor lysosomal motility in the presence and absence of rhoDV26G we made use of a green fluorescent protein-expressing vector cloned into pGEM1 harbouring the T7 promoter. Co-transfection of this plasmid with T7-myc-rhoDV26G allowed us to identify the transfected cells for video microscopy. Handling of the cells was as outlined above with the exception that they were incubated in the presence of 50 nM LysoTracker (Molecular Probes) for 15min at 37°C, 5% CO₂ to label the lysosomes, and TLVM was carried out as outlined below.

BBCE cell motility

To address the effect of rhoDV26G on cell motility, BBCE cells were trypsinized and plated at a density of 1,500 cells per cm², 24h later nuclear microinjection was carried out with a RhoDV26G expression construct at 50µg/ml concentration, FITC dextran was coinjected to identify the injected cells. Approximately, 12h following injection the cells were stimulated with bFGF and subjected to TLVM.

Time-Lapse Video Microscopy

The cells for TLVM were grown on coverslips and mounted into aluminum chambers as described in (Bradke and Dotti, 1997). Briefly, aluminum was cut into 7.5cm x 2.5cm x 2mm rectangles, the dimensions are similar to a standard glass slide and fit onto the microscope stage. In the centre a circular hole was cut of 8mm diameter, and around it 4mm area was milled from both sides. A glass coverslip was inserted onto one side of the chamber, sealed with a lubricant, medium was then added to fill the chamber, cells grown on coverslips were then inverted onto the other side of the metal slide and sealed with lubricant.

Image and statistical analysis of the data

A dedicated automatic program was developed to detect and track endosomes/lysosomes as they move. It runs on a SPARC station Ultra1 (SUN, Mountain View, CA) to which a Series 151/40 digital image processor (Imaging Technology, Bedford, MA) is connected. The detection of fluorescent spots corresponding to endosomes/lysosomes is performed automatically by a multiresolution algorithm based upon selectively filtering an undecimated wavelet decomposition of the image through the use of wavelet coefficient thresholding and correlation. At the end of this step, all endosomes/lysosomes in the sequence are characterised and their coordinates are determined and stored. A tracking algorithm is then used to establish valid trajectories. The algorithm uses a first-order Kalman filtering approach whereby at each frame and on the basis of the previous ones, predictions of the endosome/lysosome positions are established and compared with the computed ones. The best matches are selected as trajectory points and tracks are finally analysed to compute the data that was used for generating values in Tables 1 and 2. For each data set, a sequence of 30 images was analysed. Speed values reported here are mean values of the speed of several endosomes/lysosomes and errors are standard deviations of these group estimated.

Example 2:

Drugs that have been found by employing the assay of the present invention and that enhance or inhibit Rab5 effector/regulator activity can be distinguished and classified on the basis of their mode of action (activities 1 to 5, see below). Accordingly, 5 classes of drugs will be introduced (and searched for) that exhibit a specific mode of action on the target molecules, i.e., on the effectors or regulators:

1. Drugs that inhibit Rab5 activation. Such drugs would in principle act on the whole Rab5 network and are therefore less specific than drugs of classes 3 and 4 because they act on a

regulator (protein or molecule) that affects the nucleotide state of Rab5 and, consequently, the ability of Rab5 to signal to the totality of Rab5 effectors rather than on a specific effector (protein or molecule).

2. Drugs that enhance Rab5 inactivation. Such drugs would in principle act on the whole Rab5 network and are less specific than drugs of classes 3 and 4 for the reason given for class 1 above.

3. Drugs that disrupt specific Rab5-effector interactions, e.g., between Rab5 and p110 FYVE.

4. Drugs that inhibit a specific Rab5 effector activity, e.g., endosome fusion, endosome-microtubule interaction, endosome motility along microtubules.

5. Drugs that inhibit cell motility.

Class 3 and 4 drugs are the most specific and are therefore preferred according to the present invention. However, drugs of the other classes, in particular of classes 1 and 2, can nevertheless be considered as similarly suitable agents to combat at least one of the above mentioned diseases.

Assay for Activity 1

The assay measures Rabex-5 dependent nucleotide exchange of Rab5. The drug could perturb Rab5-Rabex-5 interaction or inhibit the catalytic activity of Rabex-5. The result expected is an inhibition of GDP/GTP exchange of Rab5. This assay is based on the assay published in (Horiuchi et al., 1997) and may well be adequately adapted for high throughput screening. It will be readily feasible, therefore, to screen thousands of compounds for their potential anti-cancer or anti-infectious disease activity/potential.

GDP/GTP exchange activity. Unless otherwise specified, the standard [³⁵S]GTPγS-binding assay was performed by the filter method (Sasaki et al., 1990) by incubating 200nM of recombinant Rab5 with 1μM of [³⁵S]GTPγS (20,000cpm/pmol) at 37°C for 10min with 5mM of MgCl₂ in the presence of 20nM recombinant Rabex-5.

Substances that are able to enhance or inhibit nucleotide exchange above or below the standard activity measured in the presence of Rabex-5, respectively, may be searched for.

Assay for Activity 2

The assay measures the GTPase Activating Protein (GAP) RN-Tre-dependent stimulation of GTP hydrolysis by Rab5. The drug could stabilise the Rab5-GAP interaction or stimulate the

catalytic activity of GAP. The result expected is an increase in the inactive-form of Rab5, i.e., an increased amount of GDP-bound Rab5. This assay is based on the following assay:

GAP assays with purified recombinant RN-Tre fusion proteins were performed by the filter method (Sasaki et al., 1990) using 2µM of substrate [γ -³²P]-GTP-loaded Rab5 in the presence of recombinant RN Tre GAP domain (100nM), at 30°C for 2min. GAP activity was expressed as the percentage of non-hydrolysed [γ -³²P]-GTP which remained bound to the filters, relative to the radioactivity at time 0. Substances that are able to increase the GTP hydrolysis activity compared with the standard activity measured in the presence of RN Tre should be searched for.

The assay may adequately be adapted for high throughput screening.

Assay for Activity 3

Drugs may be screened for their ability to inhibit specific Rab5-Rab5 effector interactions. The assay consists in the binding of an effector molecule (either *in vitro* translated or as purified recombinant protein; native or modified, e.g., by biotinylation) to GST-Rab5:GTPγS (GTPgammaS) immobilised on a matrix. This assay is based on the procedure developed by Christoforidis et al. (1999a and b), but may be readily adapted for high throughput screening.

GST-Rab5 affinity chromatography is performed as described previously (Christoforidis et al., 1999a). Beads with GST-Rab5 were incubated with the nucleotide exchange buffer (NE buffer) containing 20mM Hepes, 100mM NaCl, 10mM EDTA, 5mM MgCl₂, 1mM DTT, 1mM GTPγS, pH7.5, for 90min at room temperature under rotation. Afterwards, NE buffer was removed and the nucleotide was stabilised with NS buffer containing 20mM Hepes, 100mM NaCl, 5mM MgCl₂, 1mM DTT, 1mM GTPγS, pH7.5, for 20min at room temperature under rotation. *In vitro* transcription-translation of ³⁵S-methionine labeled proteins is performed according to Manufacturer's instructions (Promega). Recombinant proteins or transcribed-translated proteins (50µl of standard reactions) are incubated for 2h at 4°C with 20µl glutathione sepharose beads coupled with GST-Rab5:GTPγS, and 150µl buffer containing 20mM Hepes, 100mM NaCl, 5mM MgCl₂, 1mM DTT, and 1mM GTPγS, respectively, such that the final volume is 220µl. The incubation is performed both in the presence and in the absence of the candidate drug. When the drug is dissolved in an organic solvent the buffer in the control reaction will be supplemented with the solvent in the same concentration and amount added in the reaction in the presence of the drug. Usually, however, the buffer/solvent dissolving the drug is the same buffer used for the

assay, i.e., the buffer containing 20mM Hepes, 100mM NaCl, 5mM MgCl₂, 1mM DTT, and 1mM GTPγS. Beads are subsequently washed 4x with buffer containing 20mM Hepes, 100mM NaCl, 5mM MgCl₂, 1mM DTT, 10μM GTPγS, 1x with the same buffer but now containing 250mM NaCl, and 1x with 20mM Hepes, 250mM NaCl, 1mM DTT. Elution of bound proteins is performed as described before (Christoforidis et al., 1999a; Christoforidis and Zerial, 1999c) and eluted proteins are loaded on SDS-PAGE gel followed by immunoblotting for recombinant proteins or autoradiography for *in vitro* translated proteins.

The assay measures the activity of drugs that are capable of disrupting the association of a specific or any Rab5 effector with Rab5 in a concentration-dependent manner under the same experimental conditions that allow the interaction. For example, a drug x will disrupt the interaction of effector X with Rab5:GTPγS in a concentration-dependent manner. This drug will then be used as inhibitor of effector X in the biological process X.

A variation of the assay is to use magnetic beads in an assay of the ELISA-type (similar to the *in vitro* endosome fusion assay described below). GST-Rab5 was biotinylated using chemical crosslinking (Pierce) and nucleotide exchange was performed. Beads with GST-Rab5 were incubated with NE buffer containing 20mM Hepes, 100mM NaCl, 10mM EDTA, 5mM MgCl₂, 1mM DTT, 1mM GTPγS, pH7.5, for 90min at room temperature under rotation. Afterwards, the NE buffer was removed and the nucleotide was stabilised with NS buffer containing 20mM Hepes, 100mM NaCl, 5mM MgCl₂, 1mM DTT, 1mM GTPγS, pH7.5, for 20min at room temperature under rotation. Beads (1μl) were subsequently incubated with 1μg recombinant purified Rab5 effector coupled to a ruthenium trisbipyridine chelate (IGEN) in 20μl NS buffer in the presence or absence of chemical drug candidates. Beads were then washed three times with 500μl NS buffer and binding of the effector to Rab5 is measured with an Origen Analyzer (IGEN) according to the manufacturer procedure. The background signal (binding to Rab5:GDP, i.e., without nucleotide exchange) is deducted. Drugs that affect the interaction between Rab5 and the Rab5 effector decrease the signal in comparison with the signal obtained when the reaction is performed in the absence of the drug.

The assay measures the activity of drugs that are capable of disrupting the association of a specific or any Rab5 effector with Rab5 in a concentration-dependent manner under the same experimental conditions that allow the interaction. For example, a drug x will disrupt the

interaction of effector X with Rab5:GTPgS in a concentration-dependent manner. This drug will then be used as inhibitor of effector X in the biological process X.

Assays for Activity 4

5 The present inventors have developed three main assays to measure Rab5 and Rab5 effector activity.

(i) In vitro fusion assay (Horiuchi et al., 1997)

This assay is intended to screen for drugs that inhibit endocytosis, clathrin coated vesicle (CCV) fusion with early endosomes, early endosome homotypic fusion and endosome trafficking in
10 tumor cells or enhance endocytic membrane fusion in bacterial parasites infected cells.

Two distinct enriched populations of early endosomes labeled with either biotinylated transferrin or a sheep α -human transferrin antibody are prepared from sHeLa (suspension HeLa) cells by
15 sucrose sedimentation (Gorvel et al., 1991). The basal fusion reaction consists of the two enriched endosome populations incubated for 45minutes at 37°C in the presence of 3mg/ml of HeLa cytosol, unlabelled transferrin and an ATP regeneration system (17.3mM creatine phosphate, 87 μ g/ml creatine kinase, and 2.2mM ATP) and in the presence or absence of the candidate drug. The fusion is quantified by incubation of the fusion mix with wash buffer (50mM
20 Tris pH7.5, 100mM NaCl, 1g/l BSA and 2% (w/v) Triton X-100) and streptavidin-coated magnetic beads (Dynal). After two washes in wash buffer the samples are incubated with a rabbit α -sheep secondary antibody coupled to a ruthenium trisbipyridine chelate (IGEN) and measured with an Origen Analyzer (IGEN). The background signal (5-10% of the fusion signal) is deducted and the data expressed as the percentage of the basal fusion reaction. The CCV-early
25 endosome fusion assay is performed in the same way with biotinylated transferrin-labeled CCV and α -transferrin antibody-labeled early endosomes.

Substances that are able to inhibit CCV-endosome or early endosome fusion below the standard fusion activity in the presence of cytosol and an ATP-regenerating system may be searched for.
30 To control that the inhibition is targeted to a Rab5 effector (by inhibiting the Rab5-Rab5 effector interaction or the Rab5 effector activity), the inhibitory effect of the drug should be rescued by increasing concentrations of that particular Rab5 effector. For this a titration curve of the drug in the presence of increasing concentrations of the Rab5 effector will be necessary.

The following two assays are intended to screen for drugs that inhibit endosome movement and cell motility in tumor cells or enhance endocytic membrane fusion in infected cells. The assays score the overall activity, but as soon as the effector molecules specifically involved in these processes are identified, it will provide the additional possibility of using Assay for activity 3.

(ii) Microtubule spin-down assay

75 to 100µg purified early endosomes are incubated at room temperature for 20min with 100µg HeLa cytosol protein in a reaction brought to a final volume of 50µl by addition of BRB-80 (80mM K-Pipes, 1mM MgCl₂, 1mM EGTA, pH6.8). 10µl of taxol-stabilized microtubules (100µg tubulin equivalent) are added, and this mix is incubated for 10min at room temperature, in the presence or absence of the candidate drug, and then overlaid on 600µl of a 35% (w/v) sucrose cushion. After sedimentation at 100,000 x g for 20min at 22°C in a TLA 100.4 rotor, the upper layer was removed, the cushion was washed with 100µl BRB-80 and then also removed. The remaining pellet was resuspended in 30µl 2x SDS-PAGE buffer plus 30µl dH₂O, and analysed by SDS-PAGE and immunoblotting.

Substances that are able to inhibit the association of early endosomes with microtubules below the standard fusion activity in the presence of cytosol and an ATP-regenerating system may be searched for. To control that the inhibition is targeted to a Rab5 effector (by inhibiting the Rab5-Rab5 effector interaction or the Rab5 effector activity), the inhibitory effect of the drug may be rescued by increasing concentrations of that particular Rab5 effector. For this a titration curve of the drug in the presence of increasing concentrations of the Rab5 effector is necessary.

(iii) *In vitro* endosome motility assay

Early endosomes and cytosol are isolated as previously described (Horiuchi et al., 1997), except that HeLa spinner culture cells were allowed to internalise rhodamine-labeled transferrin for 10min at 37°C in order to label early endosomes. Oregon-green labeled tubulin is polymerized *in vitro*, and the resulting microtubules isolated, stabilised with taxol, and perfused into a microscope slide/glass coverslip chamber as previously described (Marlowe et al., 1998). The chamber is then washed with BRB-80 plus 10µM taxol to stabilise microtubules and remove non-polymerized tubulin. A mixture of HeLa cytosol (2mg/ml), MgATP (200µM), fluorescently-labeled endosomes (3mg/ml), purified bovine haemoglobin (3mg/ml; Sigma), and an anti-fade system (Howard and Hyman, 1993), in the presence or absence of the candidate drug, is perfused

into the chamber, and images of fluorescently-labeled microtubules (FITC filter set) or endosomes (Rhodamine filter set) are collected at 2sec-intervals using a time-lapse fluorescence videomicroscope. All image acquisition, processing, and analysis of movies is performed using the NIH Image v1.60 software package. Endosome movements are defined as linear, vectorial motions that occur on fluorescent microtubules over four or more consecutive images. These are discriminated from tethered Brownian motions, which generally display non-linear shaking, or flip randomly back and forth from image to image. Movements are counted from at least three and in most cases many more, individual movies, averaged and population significant differences calculated using the Excel spreadsheet program (Microsoft, Inc.). Substances that are able to inhibit the motility of early endosomes with microtubules below the standard fusion activity in the presence of cytosol and an ATP-regenerating system can be searched for. To control that the inhibition is targeted to a Rab5 effector (by inhibiting the Rab5-Rab5 effector interaction or the Rab5 effector activity), the inhibitory effect of the drug may be rescued by increasing concentrations of that particular Rab5 effector. For this a titration curve of the drug in the presence of increasing concentration of the Rab5 effector is necessary.

Assay for activity 5

This assay describes the measurement of cell motility in vitro. The assay may be standardised for high throughput screening.

To address the effect of drugs on cell motility, BBCE cells or other tumor cell lines are trypsinized and plated at a density of 1,500 cells per cm². 24h later the cells are stimulated with bFGF or other growth factors and subjected to Time-Lapse Video Microscopy (TLVM). The cells for TLVM are grown on coverslips and mounted into aluminum chambers as described in (Bradke and Dotti, 1997). Briefly, aluminum is cut into 7.5cm x 2.5cm x 2mm rectangles, the dimensions are similar to a standard glass slide and fit onto the microscope stage. In the centre a circular hole is cut of 8 mm diameter, and around it 4mm area is milled from both sides. A glass coverslip is inserted onto one side of the chamber, sealed with a lubricant, medium is then added to fill the chamber, cells grown on coverslips are then inverted onto the other side of the metal slide and sealed with lubricant. TLVM is carried out using a standard video microscopy set up. A dedicated automatic program was developed to detect and track fluorescently labeled cells as they move.

Example 3:

A novel Procedure for the Purification of Rab5 effectors.

120l of bacterial DH5 α cells were grown to express GST-Rab5 for the large scale isolation of Rab5 effectors. The cells were induced, harvested, and broken according to manufacturer instructions (Pharmacia). Lysis buffer (2500ml) consisted of a PBS solution containing 100 μ M GDP, 5mM MgCl₂, 5mM 2-mercaptoethanol, 5 μ g/ml DNAase, 5 μ g/ml RNase, and a cocktail of protease inhibitors. High speed supernatant of the cell lysate was incubated with 20ml glutathione sepharose beads (Pharmacia) at 4°C for 2h under rotation. Subsequently, beads were loaded on a column and washed with lysis buffer without DNAase, RNase, and protease inhibitors. This procedure resulted in 1g of GST-Rab5 bound to the column. The molecule is recovered in the GDP-bound inactive form and nucleotide exchange is necessary to convert it into the GTP-bound form to allow effector binding.

The present inventors have designed an alternative procedure that overcomes the low efficiency normally affecting this step. The modification consists in the use of aluminum fluoride (AlF₃) which when bound to Ras:GDP and Mg²⁺ has been shown to mimic the transition state for GTP-hydrolysis of the GTPase complexed to GAP. The inventors have obtained evidence, however, that the method can also be applied for the complex between Rab5:GDP:Mg²⁺:AlF₃ and its effectors. In other words, it is possible to obtain a complex consisting of Rab5, GDP, Mg²⁺, AlF₃, and Rab5 effectors. This procedure has for example allowed the inventors to purify at large scale EEA1 and Rabankyrin-5, suggesting that this method can be applied for the identification of novel effectors and regulators for small GTPases.

Example 4:

Construction and expression of EGFP-Rab5

pEGFP-Rab5 was constructed by PCR amplification of the human Rab5a cDNA from pGEM-myc-Rab5 using the primers CCCAAGCTTATGGCTAGTCGAGGCGCAACA and AACTGCAGTTAGTTACTACAACACTGATT followed by cloning of the HindIII-PstI fragment from the PCR product into a pEGFP-C3 expression vector (Clontech, Inc.). A431 cells were grown to ~80% confluency in 10cm petri dishes and transfected with 20-30 μ g of plasmid DNA using a calcium phosphate based protocol. Stably transfected clonal lines were isolated after incubation in selective (G418; 0.5 μ g/ml) growth medium for 7-10d and checked for GFP

fluorescence on endosomal structures. Despite the presence of the selectable marker, variable levels of expression of EGFP-Rab5 in these cells were observed.

Time-lapse fluorescence videomicroscopy of EGFP-Rab5 *in vivo*

Cells were grown on glass coverslips and were transferred to custom-built aluminum microscope slide chambers (EMBL workshop, Heidelberg) just prior to observation. Unless otherwise stated, cells expressing average levels of EGFP-Rab5 were selected and analyzed on a Zeiss Axioskop microscope using 100X/1.40 plan-Apochromat lens with a temperature-controlled objective sleeve attached (EMBL workshop, Heidelberg). Time-lapse imaging was performed, collecting images at 2sec-intervals using a computer-controlled shutter (Uniblitz, Inc.) with illumination by a 100W mercury arc lamp attenuated with two heat reflection filters and a KG-1 heat absorbance filter (Zeiss). GFP-fluorescence was visualised with Hi-Q FIT, or GFP filter sets (Chroma Technologies, Inc.), and images were acquired using a COHU 4913 CCIR video camera with on-chip integration controlled by the NIH-Image v1.60 software package.

Example 5:

Microtubule spin-down assay

75 to 100µg of purified early endosomes were incubated at room temperature for 20min with 100µg HeLa cytosol protein in a reaction brought to a final volume of 50µl by addition of BRB-80 (80mM K-Pipes, 1mM MgCl₂, 1mM EGTA, pH6.8). 10µl of taxol-stabilized microtubules (100µg tubulin equivalent) was added and this mix was incubated for 10min at room temperature, and then overlayed on 600µl of a 35% (w/v) sucrose cushion. After sedimentation at 100,000 x g for 20min at 22°C in a TLA 100.4 rotor, the upper layer was removed, the cushion was washed with 100µl BRB-80 and then also removed. The remaining pellet was resuspended in 30µl 2x SDS-PAGE buffer plus 30µl dH₂O, and analysed by SDS-PAGE and immunoblotting.

Example 6:

In vitro endosome motility assays

Early endosomes, and cytosol were isolated as previously described, except HeLa spinner culture cells were allowed to internalise rhodamine-labeled transferrin for 10min at 37°C in order to label early endosomes. Oregon-green labeled tubulin was polymerized *in vitro*, and the resulting microtubules isolated, stabilised with taxol, and perfused into a microscope slide/glass coverslip chamber as previously described. Alternatively, purified centrosomes were allowed to stick to the

glass surfaces of the chamber. Microtubule asters were then polymerized *in situ* by 30min incubation of the chamber at 37°C with fluorescently-labeled tubulin (4mg/ml) and 1mM GTP in BRB-80. The chamber was then washed with BRB-80 plus 10µM taxol to stabilise microtubules and remove non-polymerized tubulin. A mixture of HeLa cytosol (2mg/ml), MgATP (200µM), fluorescently-labeled endosomes (3mg/ml), purified bovine haemoglobin (3mg/ml; Sigma), and an anti-fade system was perfused into the chamber, and images of fluorescently-labeled microtubules (FITC filter set) or endosomes (Rhodamine filter set) were collected at 2sec-intervals using the time-lapse fluorescence videomicroscope setup described above.

10 Analysis and quantification of videos

All image acquisition, processing, and analysis of movies was performed using the NIH Image v1.60 software package. Endosome movements were defined as linear, vectorial motions that occurred on fluorescent microtubules over four or more consecutive images. These were discriminated from tethered Brownian motions, which generally displayed non-linear shaking, or flipped randomly back and forth from image to image. Movements were counted from at least three and in most cases many more, individual movies, averaged and population significant differences calculated using the Excel spreadsheet program (Microsoft, Inc.).

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CLAIMS

1. Use of an effector of a GTPase as a target in an *in vitro* or *in vivo* assay to detect
5 substances useful as pharmaceutical agents for the prophylaxis and/or treatment of cancer
and other proliferative, invasive or cell migration disorders such as endometriosis,
atherosclerosis, inflammatory and allergic diseases, infectious diseases, diabetes,
Alzheimer's disease and skin repair diseases such as psoriasis.
2. Use of claim 1, wherein the GTPase is of the Rab family.
- 10 3. Use of claim 2, wherein the GTPase is any of Rab4, Rab5, Rab7, Rab11, Rab17, Rab18,
and Rab22.
4. Use of any of the preceding claims, wherein the infectious disease is AIDS, tuberculosis,
15 pseudotuberculosis, cholera, malaria, gastroenteritis, enteric fever, typhus, those diseases
caused by pathogens (bacteria or organisms) such as Mycobacterium, Staphylococcus,
Toxoplasma, Trypanosoma, Listeria, Salmonella, Legionella, Leishmania, Coxiella,
Shigella, Yersinia, Neisseria, Vibrio, Bartonella, or any other infectious disease caused by
an infectious agent that infects cells by the endocytic route and resides intracellularly in
20 phagosomes escaping the cellular killing mechanisms.
5. Use of any of claims 1 to 3, wherein the cancer is a benign tumor, a malignant tumor, a
carcinoma, a sarcoma, a melanoma, a leukemia, a glioma, or a neuroblastoma, in
particular a lung carcinoma, an osteosarcoma, a lymphoma, a soft tissue sarcoma, a breast
25 carcinoma, a bile cancer, a cervix carcinoma, a cancer of the (small) intestine, of the
kidneys, of the cavity of the mouth, a penis carcinoma, an ovary cancer, a stomach cancer,
a cancer of the tongue, a brain cancer, a bladder carcinoma, a prostate carcinoma, a liver
carcinoma, a carcinoma of the pancreas, and every tumor that invades other tissues and
organs distinct from its site of origin.
- 30 6. Use of any of the preceding claims, wherein the assay is carried out in the presence of one
or more GTPase effector/regulator molecule(s) which is/are either native and
biochemically purified from a vertebrate, or recombinant and biochemically purified from

bacterial cultures, from yeast cultures, or from other cultured eukaryotic cells, in either case labeled by a covalent modification or radioactivity suitable for use in the assay.

7. Use of claim 6, wherein the assay is carried out in the simultaneous presence of at least one type of GTPase and/or endosomal membrane fractions fluorescently labeled or labeled by any other modification that allows its detection, and/or cytosolic extracts, and/or an ATP-regenerating system and/or a number of chemicals to be tested for their suitability as an anti-cancer or anti-infectious diseases drug.
8. Use of any of the preceding claims, wherein the substance useful as pharmaceutical agent is a molecule/substance that carries one or more of the following functional groups: a halide atom bound to an alkyl, alkenyl, alkynyl or aryl residue, an alcohol group (primary, secondary, tertiary), an ether group, a carbonyl function (aldehyde or ketone), a carboxylic acid group, a carboxylic anhydride group, a carbamoyl group, a haloformyl group, a cyano group, an ester group including a lactone group, a benzyl, phenyl, tolyl, tosyl, sulfonyl group, an amino group (primary, secondary, tertiary), an isocyanate, a cyanate, a thioisocyanate, a thiocyanate, a carbamate, an azide, a diazo group, and a quinone group; or is an organometallic compound, a sterol moiety(ies)-containing molecule, a β -hydroxy carboxylic acid, an inorganic acid or complex such as a metallocene, a nucleic acid, a cytokine, a hormone, an antibody, or an oligopeptide comprising up to 20, preferably 8, 10, or 12, amino acid residues.
9. Use of claim 8, wherein the antibody is a polyclonal or monoclonal antibody, or a fragment thereof, humanised or human, inhibitory or stimulatory, raised against and targeted towards any of the aforementioned GTPase effectors.
10. Use of claim 8, wherein the nucleic acid is genomic DNA, cDNA, or mRNA, or a fragment there, an oligonucleotide, an oligoribonucleotide, all being based on or derived from any of the GTPase effector having any of the sequences as depicted in SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, and 15, or gene therapy vectors derived from the aforementioned GTPase effector gene sequences.

11. A kit useful for carrying out the assay of any of the preceding claims, the kit comprising in a suitable means at least one GTPase effector/regulator molecule which is either native and biochemically purified from a vertebrate, or recombinant and biochemically purified from bacterial cultures, from yeast cultures, or from other cultured eukaryotic cells, in either case labeled by a covalent modification or by radioactivity suitable for the use in the assay.
12. The kit of claim 11, wherein the kit further comprises at least one type of GTPase and/or endosomal membrane fractions fluorescently labeled or labeled by any other modification that allows its detection, and/or cytosolic extracts, and/or an ATP-regenerating system and/or a number of chemicals to be tested for their suitability as a drug effective against any of the diseases depicted in claims 1, 4, and 5.
13. The kit of claim 12, wherein the at least one GTPase is/are one or more of the GTPases Rab4, Rab5, Rab7, Rab11, Rab17, Rab18, and Rab22.

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(54) Title: ASSAY TO DETECT SUBSTANCES USEFUL FOR THERAPY

(57) Abstract: The present invention relates to the use of effectors/regulators for Rab and Rho GTPases in *in vitro* and *in vivo* assays that recapitulate and measure the role of these effectors/regulators in membrane transport and membrane-cytoskeleton interactions in the endocytic pathway as novel targets to find therapeutic drugs to prevent or inhibit cancer cell growth and arrest cancer cell invasiveness as well as for stimulating and/or restoring endocytic transport and phagosome maturation in cells infected by intracellular parasites, which drugs are therefore useful for the therapy and optionally also the prophylaxis of cancer and infectious diseases. In addition, the present invention is also directed to kits useful as a means to detect drugs suitable as anti-cancer and anti-infectious diseases drugs.



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DECLARATION

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or the below named inventors are the original, first and joint inventors (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled **ASSAY TO DETECT SUBSTANCES USEFUL FOR THERAPY**, the Specification of which:

- ☐ is attached hereto.
☒ was filed on **March 15, 2002** as Application Serial No. **10/088,549**.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims.

I acknowledge the duty to disclose to the Patent and Trademark Office all information known to me to be material to patentability of the subject matter claimed in this application, as "materiality" is defined in Title 37, Code of Federal Regulations, § 1.56.

I hereby claim priority benefits under Title 35, United States Code, § 119 of any foreign application(s) for patent, United States provisional application(s), or inventor's certificate listed below and have also identified below any foreign application for patent, United States provisional application, or inventor's certificate having a filing date before that of the application on which priority is claimed:

PRIORITY APPLICATION(S)			Priority Claimed
EP 99118385.6	Europe	September 16, 1999	YES
(Number)	(Country)	(Date Filed)	Yes/No
(Number)	(Country)	(Date Filed)	Yes/No

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) listed below or any PCT international application(s) designating the United States listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application or PCT international application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose all information known to me to be material to patentability of the subject matter claimed in this application, as "materiality" is defined in Title 37, Code of Federal Regulations, § 1.56,

which become available between the filing date of the prior application and the national or PCT international filing date of this application:

PCT/EP00/09130	September 18, 2000	Pending
(Application Serial No.)	(Filing Date)	(Status)
(Application Serial No.)	(Filing Date)	(Status)

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I HEREBY DECLARE THAT ALL STATEMENTS MADE OF MY OWN KNOWLEDGE ARE TRUE AND THAT ALL STATEMENTS MADE ON INFORMATION AND BELIEF ARE BELIEVED TO BE TRUE; AND FURTHER THAT THESE STATEMENTS WERE MADE WITH THE KNOWLEDGE THAT WILLFUL FALSE STATEMENTS AND THE LIKE SO MADE ARE PUNISHABLE BY FINE OR IMPRISONMENT, OR BOTH, UNDER SECTION 1001 OF TITLE 18 OF THE UNITED STATES CODE AND THAT SUCH WILLFUL FALSE STATEMENTS MAY JEOPARDIZE THE VALIDITY OF THE APPLICATION OR ANY PATENT ISSUED THEREON.

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Arg Cys Ala Leu Leu Ala Ala Gln Ala Asn Lys Glu Ser Ser Ser Glu
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Ser Phe Ile Ser Arg Leu Leu Ala Ile Val Ala Asp Leu Tyr Glu Gln
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Glu Gln Tyr Ser Asp Leu Lys Ile Lys Val Gly Asp Arg His Ile Ser
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 Ala His Lys Phe Val Leu Ala Ala Arg Ser Asp Ser Trp Ser Leu Ala
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 Asn Leu Ser Ser Thr Lys Glu Leu Asp Leu Ser Asp Ala Asn Pro Glu
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 Val Thr Met Thr Met Leu Arg Trp Ile Tyr Thr Asp Glu Leu Glu Phe
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 Arg Glu Asp Asp Val Phe Leu Thr Glu Leu Met Lys Leu Ala Asn Arg
 130 135 140
 Phe Gln Leu Gln Leu Leu Arg Glu Arg Cys Glu Lys Gly Val Met Ser
 145 150 155 160
 Leu Val Asn Val Arg Asn Cys Ile Arg Phe Tyr Gln Thr Ala Glu Glu
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 Leu Asn Ala Ser Thr Leu Met Asn Tyr Cys Ala Glu Ile Ile Ala Ser
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 His Trp Asp Asp Leu Arg Lys Glu Asp Phe Ser Ser Met Ser Ala Gln
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 Leu Leu Tyr Lys Met Ile Lys Ser Lys Thr Glu Tyr Pro Leu His Lys
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 Ala Ile Lys Val Glu Arg Glu Asp Val Val Phe Leu Tyr Leu Ile Glu
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 Met Asp Ser Gln Leu Pro Gly Lys Leu Asn Glu Ala Asp His Asn Gly
 245 250 255
 Asp Leu Ala Leu Asp Leu Ala Leu Ser Arg Arg Leu Glu Ser Ile Ala
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 Thr Thr Leu Val Ser His Lys Ala Asp Val Asp Met Val Asp Lys Ser
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 Gly Trp Ser Leu Leu His Lys Gly Ile Gln Arg Gly Asp Leu Phe Ala
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 Ala Thr Phe Leu Ile Lys Asn Gly Ala Phe Val Asn Ala Ala Thr Leu
 305 310 315 320
 Gly Ala Gln Glu Thr Pro Leu His Leu Val Ala Leu Tyr Ser Ser Lys
 325 330 335
 Lys His Ser Ala Asp Val Met Ser Glu Met Ala Gln Ile Ala Glu Ala
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Leu Leu Gln Ala Gly Ala Asn Pro Asn Met Gln Asp Ser Lys Gly Arg
355 360 365

Thr Pro Leu His Val Ser Ile Met Ala Gly Asn Glu Tyr Val Phe Ser
370 375 380

Gln Leu Leu Gln Cys Lys Gln Leu Asp Leu Glu Leu Lys Asp His Glu
385 390 395 400

Gly Ser Thr Ala Leu Trp Leu Ala Val Gln His Ile Thr Val Ser Ser
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Asp Gln Ser Val Asn Pro Phe Glu Asp Val Pro Val Val Asn Gly Thr
420 425 430

Ser Phe Asp Glu Asn Ser Phe Ala Ala Arg Leu Ile Gln Arg Gly Ser
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His Thr Asp Ala Pro Asp Thr Ala Thr Gly Asn Cys Leu Leu Gln Arg
450 455 460

Ala Ala Gly Ala Gly Asn Glu Ala Ala Ala Leu Phe Leu Ala Thr Asn
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Gly Ala His Val Asn His Arg Asn Lys Trp Gly Glu Thr Pro Leu His
485 490 495

Thr Ala Cys Arg His Gly Leu Ala Asn Leu Thr Ala Glu Leu Leu Gln
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Gln Gly Ala Asn Pro Asn Leu Gln Thr Glu Glu Ala Leu Pro Leu Pro
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Lys Glu Ala Ala Ser Leu Thr Ser Leu Ala Asp Ser Val His Leu Gln
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Thr Pro Leu His Met Ala Ile Ala Tyr Asn His Pro Asp Val Val Ser
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Val Ile Leu Glu Gln Lys Ala Asn Ala Leu His Ala Thr Asn Asn Leu
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Gln Ile Ile Pro Asp Phe Ser Leu Lys Asp Ser Arg Asp Gln Thr Val
580 585 590

Leu Gly Leu Ala Leu Trp Thr Gly Met His Thr Ile Ala Ala Gln Leu
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Leu Gly Ser Gly Ala Ala Ile Asn Asp Thr Met Ser Asp Gly Gln Thr
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Leu Leu His Met Ala Ile Gln Arg Gln Asp Ser Lys Ser Ala Leu Phe
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Leu Leu Glu His Gln Ala Asp Ile Asn Val Arg Thr Gln Asp Gly Glu
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Asn Gly Val Asp Phe Ala Ala Val Asp Glu Asn Gly Asn Asn Ala Leu
965 970 975

His Leu Ala Val Met His Gly Arg Leu Asn Asn Ile Arg Val Leu Leu
980 985 990

Thr Glu Cys Thr Val Asp Ala Glu Ala Phe Asn Leu Arg Gly Gln Ser
995 1000 1005

Pro Leu His Ile Leu Gly Gln Tyr Gly Lys Glu Asn Ala Ala Ala Ile
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Phe Asp Leu Phe Leu Glu Cys Met Pro Gly Tyr Pro Leu Asp Lys Pro
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Asp Ala Asp Gly Ser Thr Val Leu Leu Leu Ala Tyr Met Lys Gly Asn
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Ala Asn Leu Cys Arg Ala Ile Val Arg Ser Gly Ala Arg Leu Gly Val
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Asn Asn Asn Gln Gly Val Asn Ile Phe Asn Tyr Gln Val Ala Thr Lys
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Gln Leu Leu Phe Arg Leu Leu Asp Met Leu Ser Lys Glu Pro Pro Trp
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Cys Asp Gly Ser Tyr Cys Tyr Glu Cys Thr Ala Arg Phe Gly Val Thr
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Thr Arg Lys His His Cys Arg His Cys Gly Arg Leu Leu Cys His Lys
1125 1130 1135

Cys Ser Thr Lys Glu Ile Pro Ile Ile Lys Phe Asp Leu Asn Lys Pro
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<211> 3390

<212> DNA

<213> Homo sapiens

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<211> 1129

<212> PRT

<213> Homo sapiens

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Glu Gln Tyr Ser Ala Asn Pro Glu Val Thr Met Thr Met Leu Arg Trp
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Ile Tyr Thr Asp Glu Leu Glu Phe Arg Glu Asp Asp Val Phe Leu Thr
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Glu Leu Met Lys Leu Ala Asn Arg Phe Gln Leu Gln Leu Leu Arg Glu
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Arg Cys Glu Lys Gly Val Met Ser Leu Val Asn Val Arg Asn Cys Ile
115 120 125

Arg Phe Tyr Gln Thr Ala Glu Glu Leu Asn Ala Ser Thr Leu Met Asn
130 135 140

Tyr Cys Ala Glu Ile Ile Ala Ser His Trp Asp Asp Leu Arg Lys Glu
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Asp Phe Ser Ser Met Ser Ala Gln Leu Leu Tyr Lys Met Ile Lys Ser
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Lys Thr Glu Tyr Pro Leu His Lys Ala Ile Lys Val Glu Arg Glu Asp
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Val Val Phe Leu Tyr Leu Ile Glu Met Asp Ser Gln Leu Pro Gly Lys
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Leu Asn Glu Ala Asp His Asn Gly Asp Leu Ala Leu Asp Leu Ala Leu
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Asp Val Asp Met Val Asp Lys Ser Gly Trp Ser Leu Leu His Lys Gly
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Ile Gln Arg Gly Asp Leu Phe Ala Ala Thr Phe Leu Ile Lys Asn Gly
260 265 270

Ala Phe Val Asn Ala Ala Thr Leu Gly Ala Gln Glu Thr Pro Leu His
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 305 310 315 320
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 Val Gln His Ile Thr Val Ser Ser Asp Gln Ser Val Asn Pro Phe Glu
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 Asp Val Pro Val Val Asn Gly Thr Ser Phe Asp Glu Asn Ser Phe Ala
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 Ala Arg Leu Ile Gln Arg Gly Ser His Thr Asp Ala Pro Asp Thr Ala
 405 410 415
 Thr Gly Asn Cys Leu Leu Gln Arg Ala Ala Gly Ala Gly Asn Glu Ala
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 Ala Ala Leu Phe Leu Ala Thr Asn Gly Ala His Val Asn His Arg Asn
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 Lys Trp Gly Glu Thr Pro Leu His Thr Ala Cys Arg His Gly Leu Ala
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 Asp Thr Met Ser Asp Gly Gln Thr Leu Leu His Met Ala Ile Gln Arg
 580 585 590

Gln Asp Ser Lys Ser Ala Leu Phe Leu Leu Glu His Gln Ala Asp Ile
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 Asp Ala Thr Cys Trp Gly Pro Gly Pro Gly Gly Cys Leu Gln Thr Leu
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 Cys Thr Ala Arg Phe Gly Val Thr Thr Arg Lys His His Cys Arg His
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 Cys Gly Arg Leu Leu Cys His Lys Cys Ser Thr Lys Glu Ile Pro Ile
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<211> 2406

<212> DNA

<213> Homo sapiens

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<212> PRT

<213> Homo sapiens

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Lys Ser Leu Val Gln Lys Ala Lys Lys Ala Lys Asp Arg Leu Leu Lys
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 Phe Ser Tyr Gly Gly Val Asp Pro Tyr Met Trp Glu Pro Gln Glu Leu
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 Gly Ala Val Arg Ser His Leu Ser Asp Phe Lys Lys His Arg Ala Ala
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 Arg Ile Asp His Tyr Val Val Glu Val Asn Lys Leu Ile Ile Arg Leu
 115 120 125
 Glu Lys Leu Thr Ala Phe Asp Arg Thr Asn Thr Glu Ser Ala Lys Ile
 130 135 140
 Arg Ala Ile Glu Lys Ser Val Val Pro Trp Val Asn Asp Gln Asp Val
 145 150 155 160
 Pro Phe Cys Pro Asp Cys Gly Asn Lys Phe Ser Ile Arg Asn Arg Arg
 165 170 175
 His His Cys Arg Leu Cys Gly Ser Ile Met Cys Lys Lys Cys Met Glu
 180 185 190
 Leu Ile Ser Leu Pro Leu Ala Asn Lys Leu Thr Ser Ala Ser Lys Glu
 195 200 205
 Ser Leu Ser Thr His Thr Ser Pro Ser Gln Ser Pro Asn Ser Val His
 210 215 220
 Gly Ser Arg Arg Gly Ser Ile Ser Ser Met Ser Ser Val Ser Ser Val
 225 230 235 240
 Leu Asp Glu Lys Asp Asp Asp Arg Ile Arg Cys Cys Thr His Cys Lys
 245 250 255
 Asp Thr Leu Leu Lys Arg Glu Gln Gln Ile Asp Glu Lys Glu His Thr
 260 265 270
 Pro Asp Ile Val Lys Leu Tyr Glu Lys Leu Arg Leu Cys Met Glu Lys
 275 280 285
 Val Asp Gln Lys Ala Pro Glu Tyr Ile Arg Met Ala Ala Ser Leu Asn
 290 295 300
 Ala Gly Glu Thr Thr Tyr Ser Leu Glu His Ala Ser Asp Leu Arg Val
 305 310 315 320
 Glu Val Gln Lys Val Tyr Glu Leu Ile Asp Ala Leu Ser Lys Lys Ile
 325 330 335
 Leu Thr Leu Gly Leu Asn Gln Asp Pro Pro Pro His Pro Ser Asn Leu
 340 345 350

Arg Leu Gln Arg Met Ile Arg Tyr Ser Ala Thr Leu Phe Val Gln Glu
355 360 365

Lys Leu Leu Gly Leu Met Ser Leu Pro Thr Lys Glu Gln Phe Glu Glu
370 375 380

Leu Lys Lys Lys Arg Lys Glu Glu Met Glu Arg Lys Arg Xaa Val Glu
385 390 395 400

Arg Gln Ala Ala Leu Glu Ser Gln Arg Arg Leu Glu Glu Arg Gln Ser
405 410 415

Gly Leu Ala Ser Arg Ala Ala Asn Gly Glu Val Ala Ser Leu Arg Arg
420 425 430

Gly Pro Ala Pro Leu Arg Lys Ala Glu Gly Trp Leu Pro Leu Ser Gly
435 440 445

Gly Gln Gly Gln Ser Glu Asp Ser Asp Pro Leu Leu Gln Gln Ile His
450 455 460

Asn Ile Thr Ser Phe Ile Arg Gln Ala Lys Ala Ala Gly Arg Met Asp
465 470 475 480

Glu Val Arg Thr Leu Gln Glu Xaa Leu Arg Gln Leu Gln Asp Glu Tyr
485 490 495

Asp Gln Gln Gln Thr Glu Lys Ala Ile Glu Leu Ser Arg Arg Gln Ala
500 505 510

Glu Glu Glu Asp Leu Gln Arg Glu Gln Leu Gln Met Leu Arg Glu Arg
515 520 525

Glu Leu Glu Arg Glu Arg Glu Gln Phe Arg Val Ala Ser Leu His Thr
530 535 540

Arg Thr Arg Ser Leu Asp Phe Arg Glu Ile Gly Pro Phe Gln Leu Glu
545 550 555 560

Pro Ser Arg Glu Pro Arg Thr His Leu Ala Tyr Ala Leu Asp Leu Gly
565 570 575

Ser Ser Pro Val Pro Ser Ser Thr Ala Pro Lys Thr Pro Ser Leu Ser
580 585 590

Ser Thr Gln Pro Thr Arg Val Trp Ser Gly Pro Pro Ala Val Gly Gln
595 600 605

Glu Arg Leu Pro Gln Ser Ser Met Pro Gln Gln His Glu Gly Pro Ser
610 615 620

Leu Asn Pro Phe Asp Glu Glu Asp Leu Ser Ser Pro Met Glu Glu Ala
625 630 635 640

Thr Thr Gly Pro Pro Ala Ala Gly Val Ser Leu Asp Pro Ser Ala Arg

	645		650		655
Ile Leu Lys Glu Tyr Asn Pro Phe Glu Glu Glu Asp Glu Glu Glu Glu	660		665		670
Ala Val Ala Gly Asn Pro Phe Ile Gln Pro Asp Ser Pro Ala Pro Asn	675		680		685
Pro Phe Ser Glu Glu Asp Glu His Pro Gln Gln Arg Leu Ser Ser Pro	690		695		700
Leu Val Pro Gly Asn Pro Phe Glu Glu Pro Thr Cys Ile Asn Pro Phe	705		710		715
					720
Glu Met Asp Ser Asp Ser Gly Pro Glu Ala Glu Glu Pro Ile Glu Glu	725		730		735
Glu Leu Leu Leu Gln Gln Ile Asp Asn Ile Lys Ala Tyr Ile Phe Asp	740		745		750
Ala Lys Gln Cys Gly Arg Leu Asp Glu Val Glu Val Leu Thr Glu Asn	755		760		765
Leu Arg Glu Leu Lys His Thr Leu Ala Lys Gln Lys Gly Gly Thr Asp	770		775		780

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 <211> 1659
 <212> DNA
 <213> Homo sapiens

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 agtcatttgc atggacgtct gatgctgctg cacagtttac cgtgctttat agaaaaagac 180
 ttaaaagaag ctctgactca gtttatagaa gaagaatccc tcagcgatta tgatagagat 240
 gctgaagcat ccctggcagc tgtgaaatca ggtgaagtag atttacatca gctggcgagt 300
 acatgggcca aagcttatgc tgagaccacg ttagagcatg caaggcctga agaaccacgc 360
 tgggatgaag attttgcaga tgtgtaccat gacttaattc attctcctgc ctctgaaact 420
 ctcttaaaatt tggaacataa ttactttggt agtatctcag aactgattgg tgaaagagat 480
 gtggagctga aaaaattacg agagagacaa ggtattgaaa tggaaaaagt catgcaggaa 540
 ttgggaaaat cactgacaga tcaagatgta aattcactgg ctgctcagca ttttgaatcc 600
 cagcaagacc tagaaaaata atggtcgaat gaattaaaac aatcaactgc catccaaaaa 660
 caagagtatc aagaatgggt aataaaaact caccaagacc taaaaaaccc caacaacagc 720
 tcccttagtg aggaaattaa agttcagcca agtcagttca gagaatctgt agaagcaatt 780
 ggaaggattt atgaggaaca gagaaagtta gaagaaagtt ttaccattca cttaggagcc 840
 cagttgaaga ccattgcataa tttgagattg ctgagagcag atatgctgga ctctgtgaag 900
 cataaaagaa atcatcgaag tgggtgtgaaa ctcatcggc tccaaacagc tctgtcactt 960
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164108 1975 11 11 11 11 11 11 11 11

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<210> 8
<211> 552
<212> PRT
<213> Homo sapiens
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 245 250 255
 Val Glu Ala Ile Gly Arg Ile Tyr Glu Glu Gln Arg Lys Leu Glu Glu
 260 265 270
 Ser Phe Thr Ile His Leu Gly Ala Gln Leu Lys Thr Met His Asn Leu
 275 280 285
 Arg Leu Leu Arg Ala Asp Met Leu Asp Phe Cys Lys His Lys Arg Asn
 290 295 300
 His Arg Ser Gly Val Lys Leu His Arg Leu Gln Thr Ala Leu Ser Leu
 305 310 315 320
 Tyr Ser Thr Ser Leu Cys Gly Leu Val Leu Leu Val Asp Asn Arg Ile
 325 330 335
 Asn Ser Tyr Ser Gly Ile Lys Arg Asp Phe Ala Thr Val Cys Gln Glu
 340 345 350
 Cys Thr Asp Phe His Phe Pro Arg Ile Glu Glu Gln Leu Glu Val Val
 355 360 365
 Gln Gln Val Val Leu Tyr Ala Arg Thr Gln Arg Arg Ser Lys Leu Lys
 370 375 380
 Glu Ser Leu Asp Ser Gly Asn Gln Asn Gly Gly Asn Asp Asp Lys Thr
 385 390 395 400
 Lys Asn Ala Glu Arg Asn Tyr Leu Asn Val Leu Pro Gly Glu Phe Tyr
 405 410 415
 Ile Thr Arg His Ser Asn Leu Ser Glu Ile His Val Ala Phe His Leu
 420 425 430
 Cys Val Asp Asp His Val Lys Ser Gly Asn Ile Thr Ala Arg Asp Pro
 435 440 445
 Ala Ile Met Gly Leu Arg Asn Ile Leu Lys Val Cys Cys Thr His Asp
 450 455 460
 Ile Thr Thr Ile Ser Ile Pro Leu Leu Leu Val His Asp Met Ser Glu
 465 470 475 480
 Glu Met Thr Ile Pro Trp Cys Leu Arg Arg Ala Glu Leu Val Phe Lys
 485 490 495
 Cys Val Lys Gly Phe Met Met Glu Met Ala Ser Trp Asp Gly Gly Ile
 500 505 510
 Ser Arg Thr Val Gln Phe Leu Val Pro Gln Ser Ile Ser Glu Glu Met
 515 520 525

Phe Tyr Gln Leu Ser Asn Met Leu Pro Gln Ile Phe Arg Val Ser Ser
 530 535 540

Thr Leu Thr Leu Thr Ser Lys His
 545 550

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 <211> 1050
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 <213> Homo sapiens

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 gctctgagcc agataaaatac aaagcttctg gcagaaatga agatgaaaaa ggatttattt 180
 cctgttgga gagaaattgc tgggaattgta ttagatgttg gaagcaagggt atcattcttt 240
 caaccagatg atgaagtagt tgggaatttg cccctggact ctgaagaccc tggactttgt 300
 gaagttgta gactacatga gcattacttg gttcataaac cagaaaagggt cacatggacg 360
 gaagcagcag gaagcattcg ggatggagtg cgtgcctata cagctctgca ttatctttct 420
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 gataagcagt gccttgaaag attcagacct cccatagccc gactgattga tgtatctaata 600
 gggaaagtgc atgttgctga aagctgtttg gaagaaacag gtggcctggg agtagatatt 660
 gtcctagatg ctggagtgag attatatagt aaagatgatg aaccagctgt aaaactacaa 720
 ctactaccac ataaacatga tatcatcaca cttcttggtg ttggaggcca ctgggtaaca 780
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 ctttgtatct taaaggatgt gatggagaag ttatcaactg gtgttttcag acctcagttg 960
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 ggaagaaaaa agcaagttgt tcaattttaa 1050

<210> 10
 <211> 349
 <212> PRT
 <213> Homo sapiens

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 20 25 30
 Val Lys Leu Gln Val Lys Ala Cys Ala Leu Ser Gln Ile Asn Thr Lys
 35 40 45
 Leu Leu Ala Glu Met Lys Met Lys Lys Asp Leu Phe Pro Val Gly Arg
 50 55 60
 Glu Ile Ala Gly Ile Val Leu Asp Val Gly Ser Lys Val Ser Phe Phe
 65 70 75 80

Gln	Pro	Asp	Asp	Glu	Val	Val	Gly	Ile	Leu	Pro	Leu	Asp	Ser	Glu	Asp	
				85					90						95	
Pro	Gly	Leu	Cys	Glu	Val	Val	Arg	Val	His	Glu	His	Tyr	Leu	Val	His	
			100					105					110			
Lys	Pro	Glu	Lys	Val	Thr	Trp	Thr	Glu	Ala	Ala	Gly	Ser	Ile	Arg	Asp	
		115					120					125				
Gly	Val	Arg	Ala	Tyr	Thr	Ala	Leu	His	Tyr	Leu	Ser	His	Leu	Ser	Pro	
		130				135						140				
Gly	Lys	Ser	Val	Leu	Ile	Met	Asp	Gly	Ala	Ser	Ala	Phe	Gly	Thr	Ile	
145					150					155					160	
Ala	Ile	Gln	Leu	Ala	His	His	Arg	Gly	Ala	Lys	Val	Ile	Ser	Thr	Ala	
				165					170						175	
Cys	Ser	Leu	Glu	Asp	Lys	Gln	Cys	Leu	Glu	Arg	Phe	Arg	Pro	Pro	Ile	
			180					185					190			
Ala	Arg	Val	Ile	Asp	Val	Ser	Asn	Gly	Lys	Val	His	Val	Ala	Glu	Ser	
		195					200						205			
Cys	Leu	Glu	Glu	Thr	Gly	Gly	Leu	Gly	Val	Asp	Ile	Val	Leu	Asp	Ala	
	210					215					220					
Gly	Val	Arg	Leu	Tyr	Ser	Lys	Asp	Asp	Glu	Pro	Ala	Val	Lys	Leu	Gln	
225					230					235					240	
Leu	Leu	Pro	His	Lys	His	Asp	Ile	Ile	Thr	Leu	Leu	Gly	Val	Gly	Gly	
				245					250					255		
His	Trp	Val	Thr	Thr	Glu	Glu	Asn	Leu	Gln	Leu	Asp	Pro	Pro	Asp	Ser	
			260					265					270			
His	Cys	Leu	Phe	Leu	Lys	Gly	Ala	Thr	Leu	Ala	Phe	Leu	Asn	Asp	Glu	
		275					280					285				
Val	Trp	Asn	Leu	Ser	Asn	Val	Gln	Gln	Gly	Lys	Tyr	Leu	Cys	Ile	Leu	
		290				295					300					
Lys	Asp	Val	Met	Glu	Lys	Leu	Ser	Thr	Gly	Val	Phe	Arg	Pro	Gln	Leu	
305					310					315					320	
Asp	Glu	Pro	Ile	Pro	Leu	Tyr	Glu	Ala	Lys	Val	Ser	Met	Glu	Ala	Val	
				325						330				335		
Gln	Lys	Asn	Gln	Gly	Arg	Lys	Lys	Gln	Val	Val	Gln	Phe				
			340					345								

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<210> 11
<211> 663
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<212> DNA

<213> Homo sapiens

<400> 11

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ctgcaggtgg ccaccctgg ggggaaagcc atggaatttg tggatgtgac tgagagcaat 180
gcacgctggg tgcaagactt ccgcctcaag gcttacgcca gccccgcaa gctcgagtcc 240
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tgggtgttcg acagctacag cctgacaggg ccctctgtgt gtgagctcgt cagggccccc 480
ggcttcgccc gcctgccgct cgtggtggag gacttcgtga aggattcggg cgctgcttc 540
agtgggcttg gggcagcacc aggtggggg agaggaggaa agcaagagga cagacctcca 600
gaagagcagc ggagggcggg tgaggatttc cccatatacc agtgtgtgtt catctaccca 660
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<210> 12

<211> 220

<212> PRT

<213> Homo sapiens

<400> 12

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Met Ala Ser Glu Arg Leu Pro Asn Arg Pro Ala Cys Leu Leu Val Ala
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Ser Gly Ala Ala Glu Gly Val Ser Ala Gln Ser Phe Leu His Cys Phe
          20           25           30

```

```

Thr Met Ala Ser Thr Ala Phe Asn Leu Gln Val Ala Thr Pro Gly Gly
  35           40           45

```

```

Lys Ala Met Glu Phe Val Asp Val Thr Glu Ser Asn Ala Arg Trp Val
  50           55           60

```

```

Gln Asp Phe Arg Leu Lys Ala Tyr Ala Ser Pro Ala Lys Leu Glu Ser
  65           70           75           80

```

```

Ile Asp Gly Ala Arg Tyr His Ala Leu Leu Ile Pro Ser Cys Pro Gly
          85           90           95

```

```

Ala Leu Thr Asp Leu Ala Ser Ser Gly Ser Leu Ala Arg Ile Leu Gln
 100           105           110

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```

His Phe His Ser Glu Ser Lys Pro Ile Cys Ala Val Gly His Gly Val
 115           120           125

```

```

Ala Ala Leu Cys Cys Ala Thr Asn Glu Asp Arg Ser Trp Val Phe Asp
 130           135           140

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```

Ser Tyr Ser Leu Thr Gly Pro Ser Val Cys Glu Leu Val Arg Ala Pro
 145           150           155           160

```

```

Gly Phe Ala Arg Leu Pro Leu Val Val Glu Asp Phe Val Lys Asp Ser
          165           170           175

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Gly Ala Cys Phe Ser Gly Leu Gly Ala Ala Pro Gly Trp Gly Arg Gly
 180 185 190

Gly Lys Gln Glu Asp Arg Pro Pro Glu Glu Gln Arg Arg Ala Gly Glu
 195 200 205

Asp Phe Pro Ile Tyr Gln Cys Val Phe Ile Tyr Pro
 210 215 220

<210> 13
 <211> 2679
 <212> DNA
 <213> Homo sapiens

<400> 13
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 cgctttcaaa tccttaaaac catcacccat ccagactct gccagtatgt ggatatttct 180
 aggggaaagc atgaacgact agtggcgtg gctgaacatt gtgaacgtag tctggaagac 240
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<210> 14
 <211> 892
 <212> PRT
 <213> Homo sapiens

<400> 14
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 20 25 30
 Pro Asn Ser Ile Lys Ile Leu Gly Arg Phe Gln Ile Leu Lys Thr Ile
 35 40 45
 Thr His Pro Arg Leu Cys Gln Tyr Val Asp Ile Ser Arg Gly Lys His
 50 55 60
 Glu Arg Leu Val Val Val Ala Glu His Cys Glu Arg Ser Leu Glu Asp
 65 70 75 80
 Leu Leu Arg Glu Arg Lys Pro Val Ser Cys Ser Thr Val Leu Cys Ile
 85 90 95
 Ala Phe Glu Val Leu Gln Gly Leu Gln Tyr Met Asn Lys His Gly Ile
 100 105 110
 Val His Arg Ala Leu Ser Pro His Asn Ile Leu Leu Asp Arg Lys Gly
 115 120 125
 His Ile Lys Leu Ala Lys Phe Gly Leu Tyr His Met Thr Ala His Gly
 130 135 140
 Asp Asp Val Asp Phe Pro Ile Gly Tyr Pro Ser Tyr Leu Ala Pro Glu
 145 150 155 160
 Val Ile Ala Gln Gly Ile Phe Lys Thr Thr Asp His Met Pro Ser Lys
 165 170 175
 Lys Pro Leu Pro Ser Gly Pro Lys Ser Asp Val Trp Ser Leu Gly Ile
 180 185 190
 Ile Leu Phe Glu Leu Cys Val Gly Arg Lys Leu Phe Gln Ser Leu Asp
 195 200 205
 Ile Ser Glu Arg Leu Lys Phe Leu Leu Thr Leu Asp Cys Val Asp Asp

210	215	220
Thr Leu Ile Val Leu Ala Glu Glu His Gly Cys Leu Asp Ile Ile Lys		
225	230	235 240
Glu Leu Pro Glu Thr Val Ile Asp Leu Leu Asn Lys Cys Leu Thr Phe		
	245	250 255
His Pro Ser Lys Arg Pro Thr Pro Asp Glu Leu Met Lys Asp Lys Val		
	260	265 270
Phe Ser Glu Val Ser Pro Leu Tyr Thr Pro Phe Thr Lys Pro Ala Ser		
	275	280 285
Leu Phe Ser Ser Ser Leu Arg Cys Ala Asp Leu Thr Leu Pro Glu Asp		
	290	295 300
Ile Ser Gln Leu Cys Lys Asp Ile Asn Asn Asp Tyr Leu Ala Glu Arg		
305	310	315 320
Ser Ile Glu Glu Val Tyr Tyr Leu Trp Cys Leu Ala Gly Gly Asp Leu		
	325	330 335
Glu Lys Glu Leu Val Asn Lys Glu Ile Ile Arg Ser Lys Pro Pro Ile		
	340	345 350
Cys Thr Leu Pro Asn Phe Leu Phe Glu Asp Gly Glu Ser Phe Gly Gln		
	355	360 365
Gly Arg Asp Arg Ser Ser Leu Leu Asp Asp Thr Thr Val Thr Leu Ser		
	370	375 380
Leu Cys Gln Leu Arg Asn Arg Leu Lys Asp Val Gly Gly Glu Ala Phe		
385	390	395 400
Tyr Pro Leu Leu Glu Asp Asp Gln Ser Asn Leu Pro His Ser Asn Ser		
	405	410 415
Asn Asn Glu Leu Ser Ala Ala Ala Met Leu Pro Leu Ile Ile Arg Glu		
	420	425 430
Lys Asp Thr Glu Tyr Gln Leu Asn Arg Ile Ile Leu Phe Asp Arg Leu		
	435	440 445
Lys Ala Tyr Pro Tyr Lys Lys Asn Gln Ile Trp Lys Glu Ala Arg Val		
	450	455 460
Asp Ile Pro Pro Leu Met Arg Gly Leu Thr Trp Ala Ala Leu Leu Gly		
465	470	475 480
Val Glu Gly Ala Ile His Ala Lys Tyr Asp Ala Ile Asp Lys Asp Thr		
	485	490 495
Pro Ile Pro Thr Asp Arg Gln Ile Glu Val Asp Ile Pro Arg Cys His		
	500	505 510

Gln Tyr Asp Glu Leu Leu Ser Ser Pro Glu Gly His Ala Lys Phe Arg
 515 520 525
 Arg Val Leu Lys Ala Trp Val Val Ser His Pro Asp Leu Val Tyr Trp
 530 535 540
 Gln Gly Leu Asp Ser Leu Cys Ala Pro Phe Leu Tyr Leu Asn Phe Asn
 545 550 555 560
 Asn Glu Ala Leu Ala Tyr Ala Cys Met Ser Ala Phe Ile Pro Lys Tyr
 565 570 575
 Leu Tyr Asn Phe Phe Leu Lys Asp Asn Ser His Val Ile Gln Glu Tyr
 580 585 590
 Leu Thr Val Phe Ser Gln Met Ile Ala Phe His Asp Pro Glu Leu Ser
 595 600 605
 Asn His Leu Asn Gln Ile Gly Phe Ile Pro Asp Leu Tyr Ala Ile Pro
 610 615 620
 Trp Phe Leu Thr Met Phe Thr His Val Phe Pro Leu His Lys Ile Phe
 625 630 635 640
 His Leu Trp Asp Thr Leu Leu Leu Gly Asn Ser Ser Phe Pro Phe Cys
 645 650 655
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SEQUENCE LISTING

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<110> Erik Nielsen
Savvas Chritophoridis
Carol Murphy
Marino Zerial
Stefano De Renzis

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<212> PRT

<213> Homo sapiens

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Arg Cys Ala Leu Leu Ala Ala Gln Ala Asn Lys Glu Ser Ser Ser Glu
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Ser Phe Ile Ser Arg Leu Leu Ala Ile Val Ala Asp Leu Tyr Glu Gln
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Glu Gln Tyr Ser Ala Asn Pro Glu Val Thr Met Thr Met Leu Arg Trp
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Ile Tyr Thr Asp Glu Leu Glu Phe Arg Glu Asp Asp Val Phe Leu Thr
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Glu Leu Met Lys Leu Ala Asn Arg Phe Gln Leu Gln Leu Leu Arg Glu
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Arg Cys Glu Lys Gly Val Met Ser Leu Val Asn Val Arg Asn Cys Ile
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Tyr Cys Ala Glu Ile Ile Ala Ser His Trp Asp Asp Leu Arg Lys Glu
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Asp Phe Ser Ser Met Ser Ala Gln Leu Leu Tyr Lys Met Ile Lys Ser
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Lys Thr Glu Tyr Pro Leu His Lys Ala Ile Lys Val Glu Arg Glu Asp
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Val Val Phe Leu Tyr Leu Ile Glu Met Asp Ser Gln Leu Pro Gly Lys
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Asp Val Asp Met Val Asp Lys Ser Gly Trp Ser Leu Leu His Lys Gly
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Ala Phe Val Asn Ala Ala Thr Leu Gly Ala Gln Glu Thr Pro Leu His
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Ala	Arg	Leu	Ile	Gln	Arg	Gly	Ser	His	Thr	Asp	Ala	Pro	Asp	Thr	Ala
405					410					415					
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Ala	Ala	Leu	Phe	Leu	Ala	Thr	Asn	Gly	Ala	His	Val	Asn	His	Arg	Asn
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Lys	Trp	Gly	Glu	Thr	Pro	Leu	His	Thr	Ala	Cys	Arg	His	Gly	Leu	Ala
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<213> Homo sapiens
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<212> PRT

<213> Homo sapiens

<400> 6

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Tyr Glu Glu Glu His Ser Gly Glu Asp Arg Asp Val Lys Gly Gln Ile
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 Gly Ser Arg Arg Gly Ser Ile Ser Ser Met Ser Ser Val Ser Ser Val
 225 230 235 240
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 245 250 255
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 275 280 285
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 305 310 315 320
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 325 330 335
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 340 345 350

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 355 360 365
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 370 375 380
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 385 390 395 400
 Arg Gln Ala Ala Leu Glu Ser Gln Arg Arg Leu Glu Glu Arg Gln Ser
 405 410 415
 Gly Leu Ala Ser Arg Ala Ala Asn Gly Glu Val Ala Ser Leu Arg Arg
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 Gly Pro Ala Pro Leu Arg Lys Ala Glu Gly Trp Leu Pro Leu Ser Gly
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 485 490 495
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 530 535 540
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 545 550 555 560
 Pro Ser Arg Glu Pro Arg Thr His Leu Ala Tyr Ala Leu Asp Leu Gly
 565 570 575
 Ser Ser Pro Val Pro Ser Ser Thr Ala Pro Lys Thr Pro Ser Leu Ser
 580 585 590
 Ser Thr Gln Pro Thr Arg Val Trp Ser Gly Pro Pro Ala Val Gly Gln
 595 600 605
 Glu Arg Leu Pro Gln Ser Ser Met Pro Gln Gln His Glu Gly Pro Ser
 610 615 620
 Leu Asn Pro Phe Asp Glu Glu Asp Leu Ser Ser Pro Met Glu Glu Ala
 625 630 635 640
 Thr Thr Gly Pro Pro Ala Ala Gly Val Ser Leu Asp Pro Ser Ala Arg

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675	680	685
Pro Phe Ser Glu Glu Asp Glu His Pro Gln Gln Arg Leu Ser Ser Pro		
690	695	700
Leu Val Pro Gly Asn Pro Phe Glu Glu Pro Thr Cys Ile Asn Pro Phe		
705	710	715
Glu Met Asp Ser Asp Ser Gly Pro Glu Ala Glu Glu Pro Ile Glu Glu		
725	730	735
Glu Leu Leu Leu Gln Gln Ile Asp Asn Ile Lys Ala Tyr Ile Phe Asp		
740	745	750
Ala Lys Gln Cys Gly Arg Leu Asp Glu Val Glu Val Leu Thr Glu Asn		
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Leu Arg Glu Leu Lys His Thr Leu Ala Lys Gln Lys Gly Gly Thr Asp		
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 Leu Thr Gln Phe Ile Glu Glu Glu Ser Leu Ser Asp Tyr Asp Arg Asp
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 Gln Leu Ala Ser Thr Trp Ala Lys Ala Tyr Ala Glu Thr Thr Leu Glu
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 115 120 125
 Tyr His Asp Leu Ile His Ser Pro Ala Ser Glu Thr Leu Leu Asn Leu
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 Glu His Asn Tyr Phe Val Ser Ile Ser Glu Leu Ile Gly Glu Arg Asp
 145 150 155 160
 Val Glu Leu Lys Lys Leu Arg Glu Arg Gln Gly Ile Glu Met Glu Lys
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 Val Met Gln Glu Leu Gly Lys Ser Leu Thr Asp Gln Asp Val Asn Ser
 180 185 190
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 Ser Asn Glu Leu Lys Gln Ser Thr Ala Ile Gln Lys Gln Glu Tyr Gln
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Glu Trp Val Ile Lys Leu His Gln Asp Leu Lys Asn Pro Asn Asn Ser
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 His Arg Ser Gly Val Lys Leu His Arg Leu Gln Thr Ala Leu Ser Leu
 305 310 315 320
 Tyr Ser Thr Ser Leu Cys Gly Leu Val Leu Leu Val Asp Asn Arg Ile
 325 330 335
 Asn Ser Tyr Ser Gly Ile Lys Arg Asp Phe Ala Thr Val Cys Gln Glu
 340 345 350
 Cys Thr Asp Phe His Phe Pro Arg Ile Glu Glu Gln Leu Glu Val Val
 355 360 365
 Gln Gln Val Val Leu Tyr Ala Arg Thr Gln Arg Arg Ser Lys Leu Lys
 370 375 380
 Glu Ser Leu Asp Ser Gly Asn Gln Asn Gly Gly Asn Asp Asp Lys Thr
 385 390 395 400
 Lys Asn Ala Glu Arg Asn Tyr Leu Asn Val Leu Pro Gly Glu Phe Tyr
 405 410 415
 Ile Thr Arg His Ser Asn Leu Ser Glu Ile His Val Ala Phe His Leu
 420 425 430
 Cys Val Asp Asp His Val Lys Ser Gly Asn Ile Thr Ala Arg Asp Pro
 435 440 445
 Ala Ile Met Gly Leu Arg Asn Ile Leu Lys Val Cys Cys Thr His Asp
 450 455 460
 Ile Thr Thr Ile Ser Ile Pro Leu Leu Leu Val His Asp Met Ser Glu
 465 470 475 480
 Glu Met Thr Ile Pro Trp Cys Leu Arg Arg Ala Glu Leu Val Phe Lys
 485 490 495
 Cys Val Lys Gly Phe Met Met Glu Met Ala Ser Trp Asp Gly Gly Ile
 500 505 510
 Ser Arg Thr Val Gln Phe Leu Val Pro Gln Ser Ile Ser Glu Glu Met
 515 520 525

Phe Tyr Gln Leu Ser Asn Met Leu Pro Gln Ile Phe Arg Val Ser Ser
 530 535 540

Thr Leu Thr Leu Thr Ser Lys His
 545 550

<210> 9
 <211> 1050
 <212> DNA
 <213> Homo sapiens

<400> 9
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 gctctgagcc agataaatac aaagcttctg gcagaaatga agatgaaaaa ggattttattt 180
 cctgttgagg gagaaattgc tgggaattgta ttagatgttg gaagcaagggt atcattctttt 240
 caaccagatg atgaagtagt tgggaattttg cccctggact ctgaagaccc tggactttgt 300
 gaagttgtta gagtacatga gcattacttg gttcataaac cagaaaagggt cacatggacg 360
 gaagcagcag gaagcattcg ggatggagtg cgtgcctata cagctctgca ttatctttct 420
 catctctctc ctggaaaatc agtgctgata atggatggag caagtgcatt tgggtacaata 480
 gctattcagt tagcacatca tagaggagcc aaagtgattt caacagcatg cagccttgaa 540
 gataagcagt gccttgaaaag attcagacct cccatagccc gagtgtattga tgtatctaata 600
 gggaaaagttc atgttgctga aagctgtttg gaagaaacag gtggcctggg agtagatatt 660
 gtcctagatg ctggagtgag attatatagt aaagatgatg aaccagctgt aaaactacaa 720
 ctactaccac ataaacatga tatcatcaca cttcttggtg ttggaggcca ctgggtaaca 780
 acagaagaaa accttcagtt ggatcctcca gatagccact gccttttcct caaggggagca 840
 acgttagctt tcctgaatga tgaagtttgg aatttgtcaa atgtacaaca gggaaaatat 900
 ctttgtatct taaaggatgt gatggagaag ttatcaactg gtgttttcag acctcagttg 960
 gatgaacca ttccactgta tgaggcaaaa gtttccatgg aagctgttca gaaaaatcaa 1020
 ggaagaaaaa agcaagttgt tcaattttta 1050

<210> 10
 <211> 349
 <212> PRT
 <213> Homo sapiens

<400> 10
 Met Lys Gly Leu Tyr Phe Gln Gln Ser Ser Thr Asp Glu Glu Ile Thr
 1 5 10 15
 Phe Val Phe Gln Glu Lys Glu Asp Leu Pro Val Thr Glu Asp Asn Phe
 20 25 30
 Val Lys Leu Gln Val Lys Ala Cys Ala Leu Ser Gln Ile Asn Thr Lys
 35 40 45
 Leu Leu Ala Glu Met Lys Met Lys Lys Asp Leu Phe Pro Val Gly Arg
 50 55 60
 Glu Ile Ala Gly Ile Val Leu Asp Val Gly Ser Lys Val Ser Phe Phe
 65 70 75 80

acaccagca agaaaacaaa gtccagtaaa ccaaagctcc tgggtggtga catcctgaat 2400
 agtgaagact ttattcgtgg tcacatttca ggaagcatca acattccatt cagtgcctgcc 2460
 ttactgcag aaggggagct taccagggc ccttacctg ctatgctcca gaacttcaa 2520
 gggaaggca ttgtcatcgt ggggcatgtg gcaaaacaca cagctgagtt tgcagctcac 2580
 cttgtgaaga tgaaatatcc aagaatctgt attctagatg gtggcattaa taaaataaag 2640
 ccaacaggcc tcctcaccat cccatctcct caaatatga 2679

<210> 14
 <211> 892
 <212> PRT
 <213> Homo sapiens

<400> 14
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 1 5 10 15
 Ser Ala Leu Pro His Asp Val Cys Gly Ser Asn Gly Leu Pro Leu Thr
 20 25 30
 Pro Asn Ser Ile Lys Ile Leu Gly Arg Phe Gln Ile Leu Lys Thr Ile
 35 40 45
 Thr His Pro Arg Leu Cys Gln Tyr Val Asp Ile Ser Arg Gly Lys His
 50 55 60
 Glu Arg Leu Val Val Val Ala Glu His Cys Glu Arg Ser Leu Glu Asp
 65 70 75 80
 Leu Leu Arg Glu Arg Lys Pro Val Ser Cys Ser Thr Val Leu Cys Ile
 85 90 95
 Ala Phe Glu Val Leu Gln Gly Leu Gln Tyr Met Asn Lys His Gly Ile
 100 105 110
 Val His Arg Ala Leu Ser Pro His Asn Ile Leu Leu Asp Arg Lys Gly
 115 120 125
 His Ile Lys Leu Ala Lys Phe Gly Leu Tyr His Met Thr Ala His Gly
 130 135 140
 Asp Asp Val Asp Phe Pro Ile Gly Tyr Pro Ser Tyr Leu Ala Pro Glu
 145 150 155 160
 Val Ile Ala Gln Gly Ile Phe Lys Thr Thr Asp His Met Pro Ser Lys
 165 170 175
 Lys Pro Leu Pro Ser Gly Pro Lys Ser Asp Val Trp Ser Leu Gly Ile
 180 185 190
 Ile Leu Phe Glu Leu Cys Val Gly Arg Lys Leu Phe Gln Ser Leu Asp
 195 200 205
 Ile Ser Glu Arg Leu Lys Phe Leu Leu Thr Leu Asp Cys Val Asp Asp

Gln Tyr Asp Glu Leu Leu Ser Ser Pro Glu Gly His Ala Lys Phe Arg
515 520 525

Arg Val Leu Lys Ala Trp Val Val Ser His Pro Asp Leu Val Tyr Trp
530 535 540

Gln Gly Leu Asp Ser Leu Cys Ala Pro Phe Leu Tyr Leu Asn Phe Asn
545 550 555 560

Asn Glu Ala Leu Ala Tyr Ala Cys Met Ser Ala Phe Ile Pro Lys Tyr
565 570 575

Leu Tyr Asn Phe Phe Leu Lys Asp Asn Ser His Val Ile Gln Glu Tyr
580 585 590

Leu Thr Val Phe Ser Gln Met Ile Ala Phe His Asp Pro Glu Leu Ser
595 600 605

Asn His Leu Asn Gln Ile Gly Phe Ile Pro Asp Leu Tyr Ala Ile Pro
610 615 620

Trp Phe Leu Thr Met Phe Thr His Val Phe Pro Leu His Lys Ile Phe
625 630 635 640

His Leu Trp Asp Thr Leu Leu Leu Gly Asn Ser Ser Phe Pro Phe Cys
645 650 655

Ile Gly Val Ala Ile Leu Gln Gln Leu Arg Asp Arg Leu Leu Ala Asn
660 665 670

Gly Phe Asn Glu Cys Ile Leu Leu Phe Ser Asp Leu Pro Glu Ile Asp
675 680 685

Ile Glu Arg Cys Val Arg Glu Ser Ile Asn Leu Phe Cys Trp Thr Pro
690 695 700

Lys Ser Ala Thr Tyr Arg Gln His Ala Gln Pro Pro Lys Pro Ser Ser
705 710 715 720

Asp Ser Ser Gly Gly Arg Ser Ser Ala Pro Tyr Phe Ser Ala Glu Cys
725 730 735

Pro Asp Pro Pro Lys Thr Asp Leu Ser Arg Glu Ser Ile Pro Leu Asn
740 745 750

Asp Leu Lys Ser Glu Val Ser Pro Arg Ile Ser Ala Glu Asp Leu Ile
755 760 765

Asp Leu Cys Glu Leu Thr Val Thr Gly His Phe Lys Thr Pro Ser Lys
770 775 780

Lys Thr Lys Ser Ser Lys Pro Lys Leu Leu Val Val Asp Ile Leu Asn
785 790 795 800

Ser Glu Asp Phe Ile Arg Gly His Ile Ser Gly Ser Ile Asn Ile Pro
805 810 815

<211> 583

<212> PRT

<213> Homo sapiens

<400> 16

Glu Glu Cys Gln Leu Gln Leu Lys Asn Leu His Glu Asp Leu Ser Gly
 1 5 10 15

Arg Leu Glu Glu Ser Leu Ser Ile Ile Asn Glu Lys Val Pro Phe Asn
 20 25 30

Asp Thr Lys Cys His Leu Tyr Asn Ala Leu Asn Val Pro Leu His Asn
 35 40 45

Arg Arg His Gln Leu Lys Met Arg Asp Ile Ala Gly Gln Ala Leu Ala
 50 55 60

Phe Val Gln Asp Leu Val Pro Ala Leu Leu Asn Phe His Thr Tyr Thr
 65 70 75 80

Glu Gln Arg Ile Gln Ile Phe Pro Val Asp Ser Ala Ile Asp Thr Ile
 85 90 95

Ser Pro Leu Asn Gln Lys Phe Ser Gln Tyr Leu His Glu Asn Ala Ser
 100 105 110

Tyr Val Arg Pro Leu Glu Glu Gly Met Leu His Leu Phe Glu Ser Ile
 115 120 125

Thr Glu Asp Thr Val Thr Val Leu Glu Thr Thr Val Lys Leu Lys Met
 130 135 140

Phe Ser Asp His Leu Thr Ser Tyr Val Arg Phe Leu Arg Lys Ile Leu
 145 150 155 160

Pro Tyr Gln Leu Lys Ser Leu Glu Glu Glu Cys Glu Ser Ser Leu Cys
 165 170 175

Thr Pro Ala Leu Arg Ala Arg Asn Leu Glu Leu Ser Gln Asp Met Lys
 180 185 190

Thr Met Thr Ala Val Phe Glu Lys Leu Gln Thr Tyr Val Thr Leu Leu
 195 200 205

Ala Leu Pro Ser Thr Glu Pro Asp Gly Leu Leu Arg Thr Asn Tyr Thr
 210 215 220

Ser Val Leu Thr Asn Val Gly Ala Ala Leu His Gly Phe His Asp Val
 225 230 235 240

Met Lys Asp Ile Ser Lys His Tyr Ser Gln Lys Ala Ser Ile Glu His
 245 250 255

Glu Ile Pro Thr Ala Thr Gln Lys Leu Val Thr Thr Asn Asp Cys Ile

260	265	270
Leu Ser Ser Ala Val Thr Leu Thr Asn Gly Ala Gly Lys Ile Ala Ser		
275	280	285
Phe Phe Gly Asn Asn Val Asp Tyr Phe Ile Ala Ser Leu Ser Tyr Gly		
290	295	300
Pro Lys Thr Ala Ser Gly Phe Ile Ser Pro Leu Ser Ala Glu Cys Met		
305	310	315
Leu Gln Tyr Lys Lys Lys Ala Ala Ala Tyr Met Lys Ser Leu Arg Thr		
325	330	335
Pro Leu Ala Glu Ser Val Pro Tyr Gly Glu Ala Val Ala Asn Arg Arg		
340	345	350
Val Leu Leu Ser Ser Thr Glu Ser Arg Glu Gly Leu Ala Gln Gln Val		
355	360	365
Gln Gln Ser Leu Glu Lys Ile Ser Lys Leu Glu Gln Glu Lys Glu His		
370	375	380
Trp Met Leu Glu Ala Gln Leu Ala Lys Ile Lys Leu Glu Lys Glu Asn		
385	390	395
Gln Arg Ile Ala Asp Arg Leu Arg Gly Thr Thr Ser Ala Gln Leu Pro		
405	410	415
Gly Leu Ala Gln Glu Asn Ala Thr Val Pro Ile Ala Ser Ser Gln Glu		
420	425	430
Glu Ala Ala Ala Lys Val Leu Thr Glu Pro Val Gln Ser Thr Ser Leu		
435	440	445
Val Gly Met Leu Thr Arg Thr Pro Asp Ser Glu Ala Pro Asp Val Glu		
450	455	460
Ser Arg Glu Asp Leu Ile Lys Ser His Tyr Met Ala Arg Ile Ala Glu		
465	470	475
Leu Thr Ser Gln Leu Gln Leu Ala Asp Ser Lys Ser Val His Phe Tyr		
485	490	495
Ala Glu Cys Arg Ala Leu Ser Lys Arg Leu Ala Leu Ala Glu Lys Ser		
500	505	510
Lys Glu Thr Leu Thr Glu Glu Met Arg Leu Ala Ser Gln Asn Ile Ser		
515	520	525
Arg Leu Gln Asp Glu Leu Met Thr Thr Lys Arg Ser Tyr Glu Asp Gln		
530	535	540
Leu Ser Met Met Ser Asp His Leu Cys Ser Met Asn Glu Thr Leu Ser		
545	550	555
		560

ALL INFORMATION CONTAINED HEREIN IS UNCLASSIFIED

Lys Gln Arg Glu Glu Ile Asp Thr Leu Lys Met Ala Ser Lys Gly Asn
 565 570 575

Ser Lys Lys Thr Arg Asn Arg
580